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**HODGKIN LYMPHOMA  
NON-HODGKIN LYMPHOMAS**

**CHISINAU – 2020**

# **WHO CLASSIFICATION OF MYELOID NEOPLASMS AND ACUTE LEUKEMIA**

(VARDIMAN et al., BLOOD, 30 JULY 2009 VOLUME 114, NUMBER 5)

## **MYELOPROLIFERATIVE NEOPLASMS (MPN)**

- Chronic myelogenous leukemia, *BCR-ABL1*–positive**
- Chronic neutrophilic leukemia**
- Polycythemia vera**
- Primary myelofibrosis**
- Essential thrombocythemia**
- Chronic eosinophilic leukemia, not otherwise specified**
- Mastocytosis**
- Myeloproliferative neoplasms, unclassifiable**

## **MYELOID AND LYMPHOID NEOPLASMS ASSOCIATED WITH EOSINOPHILIA AND ABNORMALITIES OF *PDGFRA*, *PDGFRB*, OR *FGFR1***

- Myeloid and lymphoid neoplasms associated with *PDGFRA* rearrangement**
- Myeloid neoplasms associated with *PDGFRB* rearrangement**
- Myeloid and lymphoid neoplasms associated with *FGFR1* abnormalities**

## **MYELOYDYSPLASTIC/MYELOPROLIFERATIVE NEOPLASMS (MDS/MPN)**

- Chronic myelomonocytic leukemia**
- Atypical chronic myeloid leukemia, *BCR-ABL1*–negative**
- Juvenile myelomonocytic leukemia**
- Myelodysplastic/myeloproliferative neoplasm, unclassifiable**
  - Provisional entity: refractory anemia with ring sideroblasts and thrombocytosis***

# **WHO CLASSIFICATION OF MYELOID NEOPLASMS AND ACUTE LEUKEMIA**

(VARDIMAN et al., BLOOD, 30 JULY 2009 VOLUME 114, NUMBER 5)

## **MYELODYSPLASTIC SYNDROME (MDS)**

**Refractory cytopenia with unilineage dysplasia**

**Refractory anemia**

**Refractory neutropenia**

**Refractory thrombocytopenia**

**Refractory anemia with ring sideroblasts**

**Refractory cytopenia with multilineage dysplasia**

**Refractory anemia with excess blasts**

**Myelodysplastic syndrome with isolated del(5q)**

**Myelodysplastic syndrome, unclassifiable**

**Childhood myelodysplastic syndrome**

*Provisional entity: refractory cytopenia of childhood*

## **ACUTE MYELOID LEUKEMIA AND RELATED NEOPLASMS**

**Acute myeloid leukemia with recurrent genetic abnormalities**

**AML with t(8;21)(q22;q22); *RUNX1-RUNX1T1***

**AML with inv(16)(p13.1q22) or t(16;16)(p13.1;q22); *CBFB-MYH11***

**APL with t(15;17)(q22;q12); *PML-RARA***

**AML with t(9;11)(p22;q23); *MLLT3-MLL***

**AML with t(6;9)(p23;q34); *DEK-NUP214***

**AML with inv(3)(q21q26.2) or t(3;3)(q21;q26.2); *RPN1-EVI1***

**AML (megakaryoblastic) with t(1;22)(p13;q13); *RBM15-MKL1***

*Provisional entity: AML with mutated NPM1*

*Provisional entity: AML with mutated CEBPA*

# **WHO CLASSIFICATION OF MYELOID NEOPLASMS AND ACUTE LEUKEMIA**

(VARDIMAN et al., BLOOD, 30 JULY 2009 VOLUME 114, NUMBER 5)

**Acute myeloid leukemia with myelodysplasia-related changes**

**Therapy-related myeloid neoplasms**

**Acute myeloid leukemia, not otherwise specified**

**AML with minimal differentiation**

**AML without maturation**

**AML with maturation**

**Acute myelomonocytic leukemia**

**Acute monoblastic/monocytic leukemia**

**Acute erythroid leukemia**

**Pure erythroid leukemia**

**Erythroleukemia, erythroid/myeloid**

**Acute megakaryoblastic leukemia**

**Acute basophilic leukemia**

**Acute panmyelosis with myelofibrosis**

**Myeloid sarcoma**

**Myeloid proliferations related to Down syndrome**

**Transient abnormal myelopoiesis**

**Myeloid leukemia associated with Down syndrome**

**Blastic plasmacytoid dendritic cell neoplasm**

# WHO CLASSIFICATION OF MYELOID NEOPLASMS AND ACUTE LEUKEMIA

(VARDIMAN et al., BLOOD, 30 JULY 2009 VOLUME 114, NUMBER 5)

## ACUTE LEUKEMIAS OF AMBIGUOUS LINEAGE

Acute undifferentiated leukemia

Mixed phenotype acute leukemia with t(9;22)(q34;q11.2); *BCR-ABL1*

Mixed phenotype acute leukemia with t(v;11q23); *MLL* rearranged

Mixed phenotype acute leukemia, B-myeloid, NOS

Mixed phenotype acute leukemia, T-myeloid, NOS

*Provisional entity: natural killer (NK) cell lymphoblastic leukemia/lymphoma*

## B LYMPHOBLASTIC LEUKEMIA/LYMPHOMA

B lymphoblastic leukemia/lymphoma, NOS

B lymphoblastic leukemia/lymphoma with recurrent genetic abnormalities

B lymphoblastic leukemia/lymphoma with t(9;22)(q34;q11.2); *BCR-ABL 1*

B lymphoblastic leukemia/lymphoma with t(v;11q23); *MLL* rearranged

B lymphoblastic leukemia/lymphoma with t(12;21)(p13;q22) *TEL-AML1*  
(*ETV6-RUNX1*)

B lymphoblastic leukemia/lymphoma with hyperdiploidy

B lymphoblastic leukemia/lymphoma with hypodiploidy

B lymphoblastic leukemia/lymphoma with t(5;14)(q31;q32) *IL3-IGH*

B lymphoblastic leukemia/lymphoma with t(1;19)(q23;p13.3); *TCF3-PBX1*

## T LYMPHOBLASTIC LEUKEMIA/LYMPHOMA

# **WHO CLASSIFICATION OF TUMOURS OF HAEMATOPOIETIC AND LYMPHOID TISSUES**

(Swerdlow S.H., Campo E, Harris NL et al, eds. Lyon: IARC Press, 2008)

## **MATURE B-CELL NEOPLASMS**

**Chronic lymphocytic leukemia/small lymphocytic lymphoma**

**B-cell prolymphocytic leukemia**

**Splenic B-cell marginal zone lymphoma**

**Hairy cell leukemia**

**Splenic lymphoma/leukemia, unclassifiable (provisional)**

**Splenic diffuse red pulp small B-cell lymphoma (provisional)**

**Hairy cell leukemia-variant (provisional)**

**Lymphoplasmacytic lymphoma**

**Waldenström macroglobulinemia**

**Heavy chain diseases**

**α Heavy chain disease**

**γ Heavy chain disease**

**μ Heavy chain disease**

**Plasma cell myeloma**

**Solitary plasmacytoma of bone**

**Extraosseous plasmacytoma**

**Extranodal marginal zone lymphoma of mucosa-associated lymphoid tissue (MALT lymphoma)**

**Nodal marginal zone lymphoma**

**Pediatric nodal marginal zone lymphoma (provisional)**

**Follicular lymphoma**

**Pediatric follicular lymphoma (provisional)**

# **WHO CLASSIFICATION OF TUMOURS OF HAEMATOPOIETIC AND LYMPHOID TISSUES**

(Swerdlow S.H., Campo E, Harris NL et al, eds. Lyon: IARC Press, 2008)

**Primary cutaneous follicle center lymphoma**

**Mantle cell lymphoma**

**Diffuse large B-cell lymphoma (DLBCL), not otherwise specified**

**T-cell/histiocyte-rich large B-cell lymphoma**

**Primary DLBCL of the CNS**

**Primary cutaneous DLBCL, leg type**

**EBV-positive DLBCL of the elderly (provisional)**

**DLBCL associated with chronic inflammation**

**Lymphomatoid granulomatosis**

**Primary mediastinal (thymic) large B-cell lymphoma**

**Intravascular large B-cell lymphoma**

**ALK-positive DLBCL**

**Plasmablastic lymphoma**

**Large B-cell lymphoma arising in HHV8-associated multicentric**

**Castleman disease**

**Primary effusion lymphoma**

**Burkitt lymphoma**

**B-cell lymphoma, unclassifiable, with features intermediate between  
diffuse large B-cell lymphoma and Burkitt lymphoma**

**B-cell lymphoma, unclassifiable, with features intermediate between  
diffuse large B-cell lymphoma and classic Hodgkin lymphoma**

# **WHO CLASSIFICATION OF TUMOURS OF HAEMATOPOIETIC AND LYMPHOID TISSUES**

(Swerdlow S.H., Campo E, Harris NL et al, eds. Lyon: IARC Press, 2008)

## **MATURE T-CELL AND NK CELL NEOPLASMS**

**T-cell prolymphocytic leukemia**

**T-cell large granular lymphocytic leukemia**

**Chronic lymphoproliferative disorder of NK cells (provisional)**

**Aggressive NK cell leukemia**

**Systemic EBV-positive T-cell lymphoproliferative disease of childhood**

**Hydroa vacciniforme-like lymphoma**

**Adult T-cell leukemia/lymphoma**

**Extranodal NK/T-cell lymphoma, nasal type**

**Enteropathy-associated T-cell lymphoma**

**Hepatosplenic T-cell lymphoma**

**Subcutaneous panniculitis-like T-cell lymphoma**

**Mycosis fungoides**

**Sézary syndrome**

**Primary cutaneous CD30+ T-cell lymphoproliferative disorders**

**Lymphomatoid papulosis**

**Primary cutaneous anaplastic large cell lymphoma**

**Primary cutaneous *g/d* T-cell lymphoma**

**Primary cutaneous CD8+ aggressive epidermotropic cytotoxic T-cell lymphoma (provisional)**

**Primary cutaneous CD4+ small/medium T-cell lymphoma (provisional)**

**Peripheral T-cell lymphoma, not otherwise specified**

**Angio-immunoblastic T-cell lymphoma**

**Anaplastic large cell lymphoma, ALK positive**

**Anaplastic large cell lymphoma, ALK negative (provisional)**



# **WHO CLASSIFICATION OF TUMOURS OF HAEMATOPOIETIC AND LYMPHOID TISSUES**

(Swerdlow S.H., Campo E, Harris NL et al, eds. Lyon: IARC Press, 2008)

## **HODGKIN LYMPHOMA**

**Nodular lymphocyte-predominant Hodgkin lymphoma**

**Classic Hodgkin lymphoma**

**Nodular sclerosis classic Hodgkin lymphoma**

**Lymphocyte-rich classic Hodgkin lymphoma**

**Mixed cellularity classic Hodgkin lymphoma**

**Lymphocyte-depleted classic Hodgkin lymphoma**

## **HISTIOCYTIC AND DENDRITIC CELL NEOPLASMS**

**Histiocytic sarcoma**

**Langerhans cell histiocytosis**

**Langerhans cell sarcoma**

**Interdigitating dendritic cell sarcoma**

**Follicular dendritic cell sarcoma**

**Fibroblastic reticular cell tumor**

**Indeterminate dendritic cell tumor**

**Disseminated juvenile xanthogranuloma**

## **POST-TRANSPLANT LYMPHOPROLIFERATIVE DISORDERS (PTLD)**

**Early lesions**

**Plasmacytic hyperplasia**

**Infectious mononucleosis-like PTLD**

**Polymorphic PTLD**

**Monomorphic PTLD (B- and T/NK cell types)**

**Classic Hodgkin lymphoma type PTLD**

## **B-CELL DEVELOPMENT AND THE CORRESPONDING LYMPHOMAS DERIVED AT EACH STAGE**

(American Society of Hematology, New Approaches to Lymphoma Diagnosis. Hematology 2001)

	B-Cells	Immunoglobulin Genes	Somatic Mutations	Ig Protein	Marker	Corresponding Lymphoma	
Foreign antigen independent	Stem cell	Germ line	None	None	CD34		Bone marrow
	Pro-B-cell §	Germ line	None	None	CD19, CD79a, BSAP, CD34, CD10, TdT		
	Pre-B-cell *	IgH rearrangement μ-chain (Cytoplasm)	None	Igμ	CD19, CD45R, CD79a, BSAP, CD34, CD10, TdT	<b>B-LBL/ALL</b>	
Foreign antigen dependent	Immature B-Cell	IgL/IgH-rearrangements IgM (Membrane)	None	IgM (Membrane)	CD19, CD20, CD45R, CD79a, CD10 BSAP		Peripheral lymphoid tissue
	Mature naive B-cell	IgH/L rearrangements IgM und IgD (Membrane)	None	IgM/IgD	CD19, CD20, CD45R, CD79a, BSAP, CD5	<b>B-CLL, MCL</b>	
	Germinal Center (CB and CC)	IgH/L rearrangements Class switch	Introduction of somatic mutations	Ig (minimal or absent)	CD19, CD20, CD45R, CD79a, BSAP, CD10, BCL6	<b>BL, FL, LPHL, DLBCL, cHL§</b>	
Terminal Differentiation	Memory B-Cell	IgH/L rearrangements	Somatic mutations	IgM	CD19, CD20, CD45R, CD79a, BSAP	<b>MZL, B-CLL</b>	
	Plasma cell	IgH/L rearrangements	Somatic mutations	IgG>IgA>IgD	CD38, Vs38c, MUM-1, CD138	<b>Plasmacytoma/ myeloma</b>	

§ There is a developmental stage between the Pro-B-cell and the Pre-B-cell for which no universally accepted term exists. Terms previously used are: "pre-pre-B" or "common B-cell precursor." This intermediate cellular stage most commonly gives rise to B-LBL/ALL.

\* For a detailed description of the Ig-gene rearrangement events early and late pre-B-cells are distinguished (see Table 2).

§ The relationship to germinal center cells can only be determined by molecular biological investigations, as the phenotype of the tumor cells is completely changed following the malignant transformation.

Abbreviations: CB, centroblasts; CC, centrocytes; Ig, Immunoglobulin; B-LBL, B-cell lymphoblastic lymphoma; B-CLL, chronic lymphocytic leukemia; MCL, mantle cell lymphoma; BL, Burkitt lymphoma; FL, follicle center lymphoma; LPHL, lymphocyte-predominant Hodgkin lymphoma; DLBCL, diffuse large cell B-cell lymphoma; cHL, classic Hodgkin lymphoma; MZL, marginal zone B-cell lymphoma.

## **HODGKIN LYMPHOMA**

**HODGKIN LYMPHOMA (HL) is a clinicopathologically aggressive B-cell lymphoma, which is one of the most curable of all haematological malignancies. The annual incidence of HL ranges between 1.47 – 2.8 cases per 100.000 population. This malignancy occurs mostly in population with the age of 5 – 9, 40 – 49, 60 – 69 years old in males and 20 – 29, 60 – 69 years old in females. Male : female ratio may reach 1.4-1.5 : 1.**

**The demonstration of clonal rearrangements of the immunoglobulin heavy- and light-chain loci has confirmed their B-cell origin. The presence of somatic hypermutation within the immunoglobulin heavy and light chain loci suggest that HRS cells are derived from B-lymphocytes with germinal centre exposure. Despite their B-cell ontogenesis, HRS cells have lost most of the normal B-cell lineage gene expression program (including the expression of immunoglobulin) through numerous aberrant genetic mechanisms, such as epigenetic silencing at B-cell gene promoter regions.**

Source: Postgraduate Haematology, Seventh Edition. Edited by A Victor Hoffbrand, Douglas R Higgs, David M Keeling and Atul B Mehta. © 2016 JohnWiley & Sons, Ltd. Published 2016 by JohnWiley & Sons, Ltd.

## HODGKIN LYMPHOMA

**MOLECULAR PATHOGENESIS.** Genes involved in normal B-lymphocyte growth and differentiation are suppressed in HRS cells. Instead, numerous aberrant intracellular signalling pathways contribute to the malignant phenotype of HRS cells including the following:

HRS cells show constitutive activation of the NF- $\kappa$ B pathway, which is associated with apoptosis resistance. The basis for constitutive NF- $\kappa$ B activation in at least a proportion of cases is the result of inactivating mutations in *TNFAIP3* and *NFKBIA*, which encode inhibitors of the NF- $\kappa$ B pathway.

The JAK-STAT signalling pathway is overactive in HRS cells, resulting in uncontrolled growth and proliferation. Mechanisms of JAK-STAT over-activity include chromosomal gains at 9p24.1-24.3 (which includes the JAK2 locus) and inactivating mutations in *PTPN1* (leading to increased phosphorylation of JAK-STAT pathway members).

HRS cells have been shown to have deacetylated histones (H3), increased H3K27 trimethylation and DNA methylation patterns, leading to silencing of tumour-suppressor genes and the extinction of the normal B-lymphocyte expression profile.

High-throughput sequencing studies have detected deleterious mutations in  $\beta$ 2M which could potentially contribute to immune evasion by HRS cells through decreased  $\beta$ 2M expression.

**CLINICAL EXAMINATION OF PATIENT WITH HODGKIN LYMPHOMA:  
CERVICAL LYMPH NODE ENLARGEMENT**

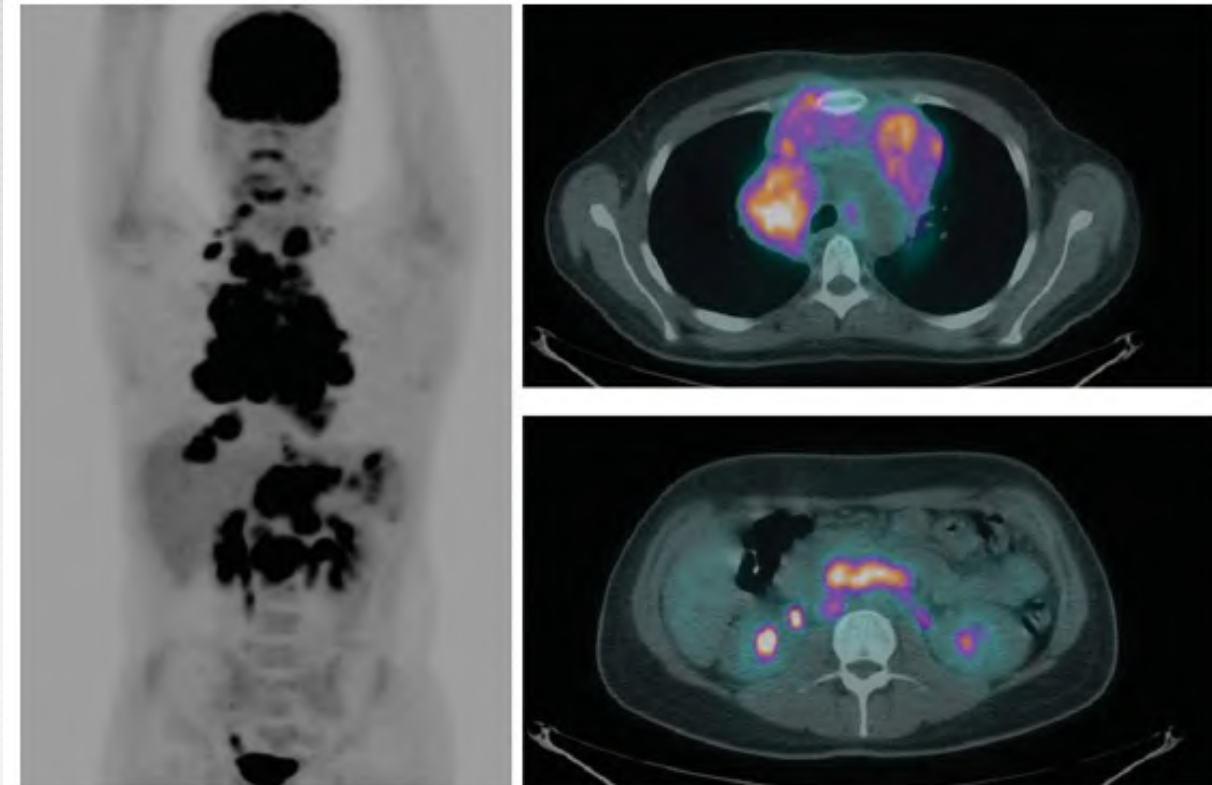


**X-RAY EXAMINATION IN HODGKIN LYMPHOMA**



**Thorax survey demonstrates mediastinal lymph node enlargement**

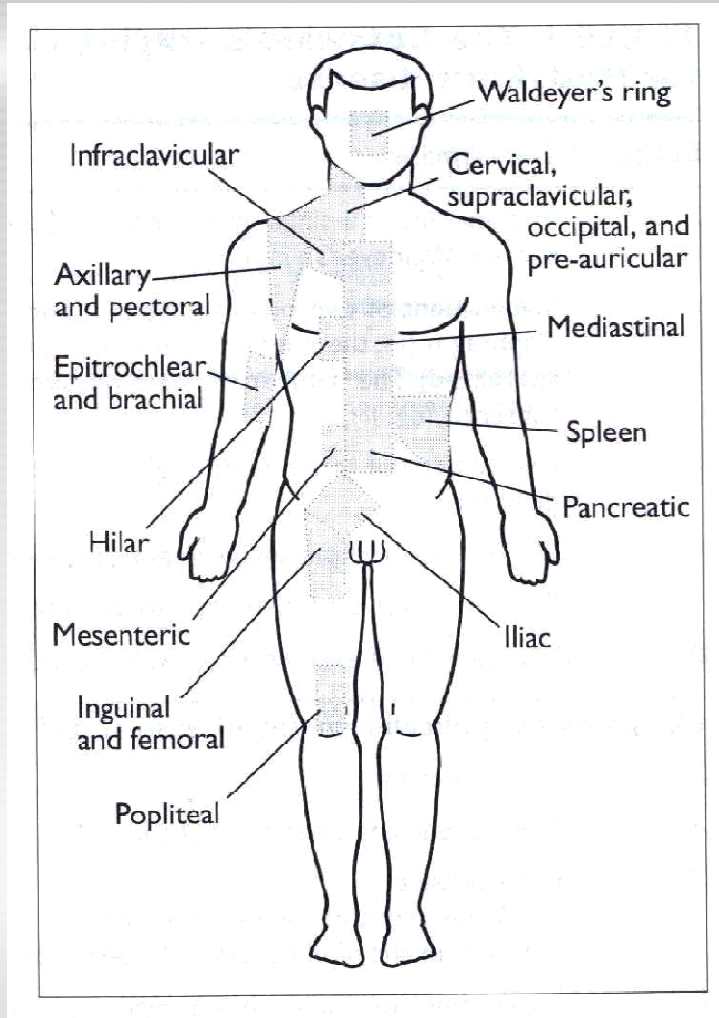
***PET scan images of patient with stage III HL showing FDG-avid cervical, mediastinal, mesenteric and retroperitoneal lymphadenopathy***



Source: Postgraduate Haematology, Seventh Edition. Edited by A Victor Hoffbrand, Douglas R Higgs, David M Keeling and Atul B Mehta. © 2016 JohnWiley & Sons, Ltd. Published 2016 by JohnWiley & Sons, Ltd.

## ANATOMIC REGIONS FOR STAGING OF HODGKIN LYMPHOMA

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach. 8th Edition. New York: CMP Healthcare Media)





## STAGING CLASSIFICATION OF HODGKIN LYMPHOMA:

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach. 8th Edition. New York: CMP Healthcare Media)

Stage	Description
I	Involvement of a single lymph node region or lymphoid structure (eg, spleen, thymus, Waldeyer's ring)
II	Involvement of two or more lymph node regions on the same side of the diaphragm (ie, the mediastinum is a single site, hilar lymph nodes are lateralized). The number of anatomic sites should be indicated by a subscript (eg, II <sub>2</sub> ).
III	Involvement of lymph node regions or structures on both sides of the diaphragm: III <sub>1</sub> : With or without involvement of splenic, hilar, celiac, or portal nodes III <sub>2</sub> : With involvement of para-aortic, iliac, or mesenteric nodes
IV	Involvement of extranodal site(s) beyond that designated E

**Designations applicable to any disease stage<sup>a</sup>**

A	No symptoms
B	Fever, drenching sweats, weight loss
X	Bulky disease: > 1/3 the width of the mediastinum > 10 cm maximal dimension of nodal mass
E	Involvement of a single extranodal site, contiguous or proximal to a known nodal site
CS	Clinical stage
PS	Pathologic stage

<sup>a</sup> For examples of how these designations are applied to disease stage, see text discussion.

**Elevated levels of fibrinogen (>5.0 g/l),  $\alpha_2$ -globulin (>10 g/l), erythrocyte sedimentation rate (> 30mm/h), haptoglobin (>1.5mg%) are considered the signs of biological activity of Hodgkin lymphoma (if present, designation *b* is used).**

## **PROGNOSTIC SCORING STAGING SYSTEM FOR HODGKIN LYMPHOMA:**

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach. 8th Edition. New York: CMP Healthcare Media)

### **EARLY STAGES**

<b>Favorable</b>	CS I and II (maximum 3 involved areas) and < 50 years <i>and</i> ESR < 50 mm/h (no B symptoms) <i>or</i> ESR < 30 mm/h (B symptoms present) <i>and</i> MT ratio < 0.35
<b>Unfavorable</b>	CS II $\geq$ 4 nodal areas involved <i>or</i> age $\geq$ 50 years <i>or</i> ESR $\geq$ 50 mm/h (no B symptoms) <i>or</i> ESR $\geq$ 30 mm/h (B symptoms present) <i>or</i> MT ratio $\geq$ 0.35

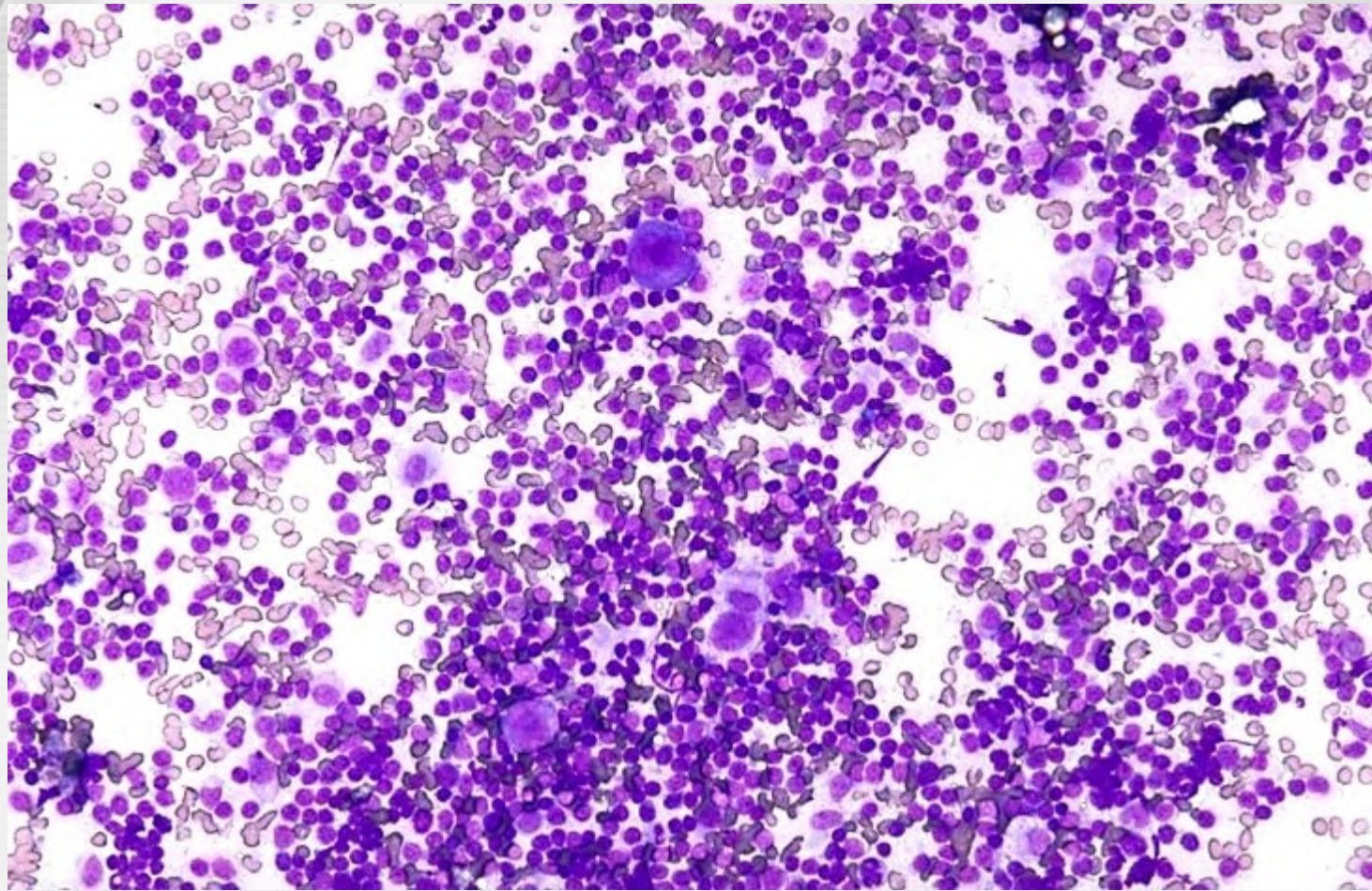
CS = Cotswald's staging; EORTC = European Organization for Research and Treatment of Cancer  
ESR = erythrocyte sedimentation rate; MT = mediastinal/thoracic

### **ADVANCED STAGES**

Serum albumin <4 g/dL  
Hemoglobin <10.5 g/dL  
Male sex  
Stage IV disease  
Age  $\geq$ 45 years  
White cell count >15,000/ $\mu$ L  
Lymphocyte count <600/ $\mu$ L *or* <8% of total white cell count

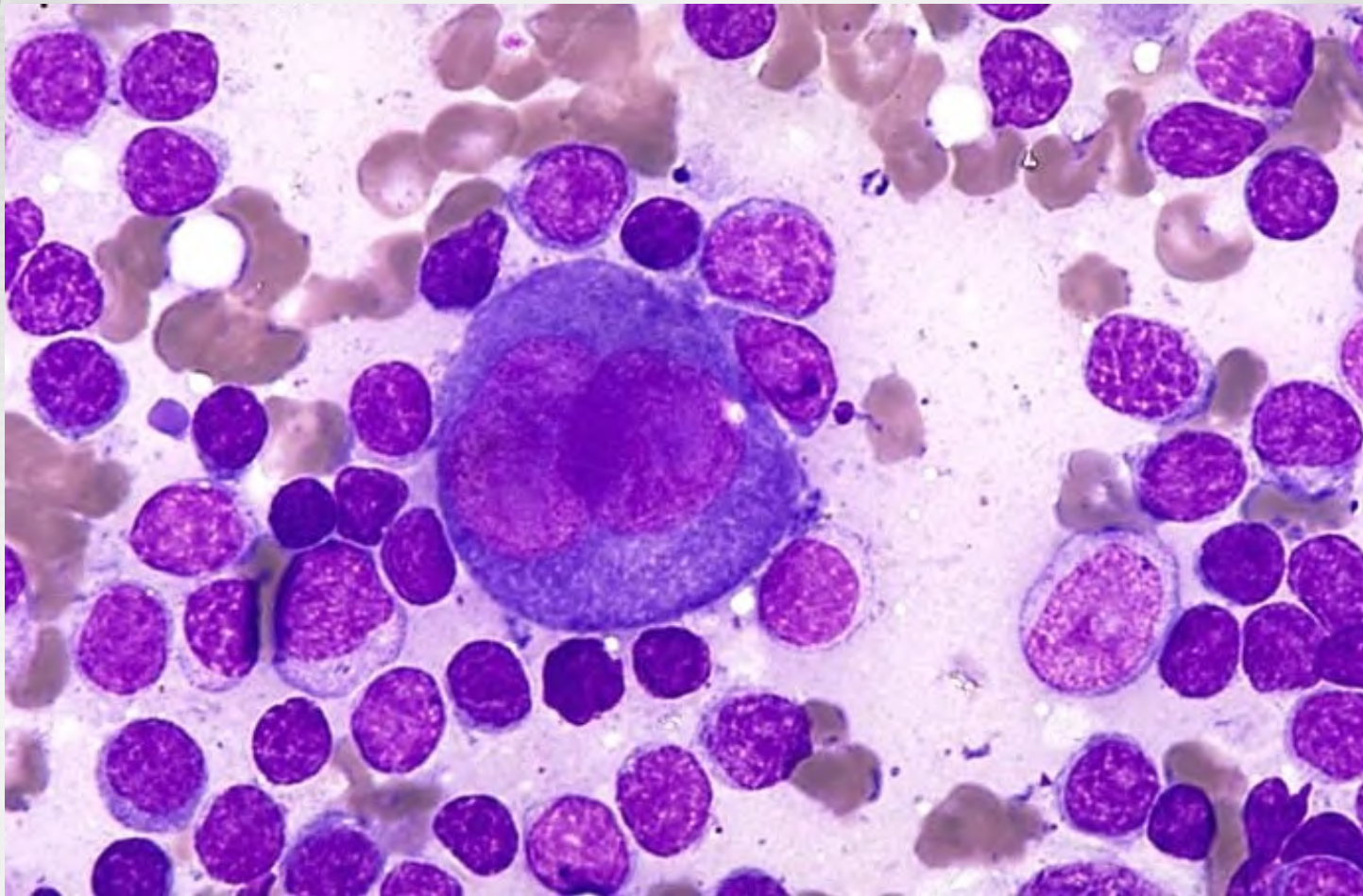
\*From Hasenclever D, Diehl V. A prognostic score for advanced Hodgkin disease. N Engl J Med. 1998;339:1506.

## HODGKIN LYMPHOMA



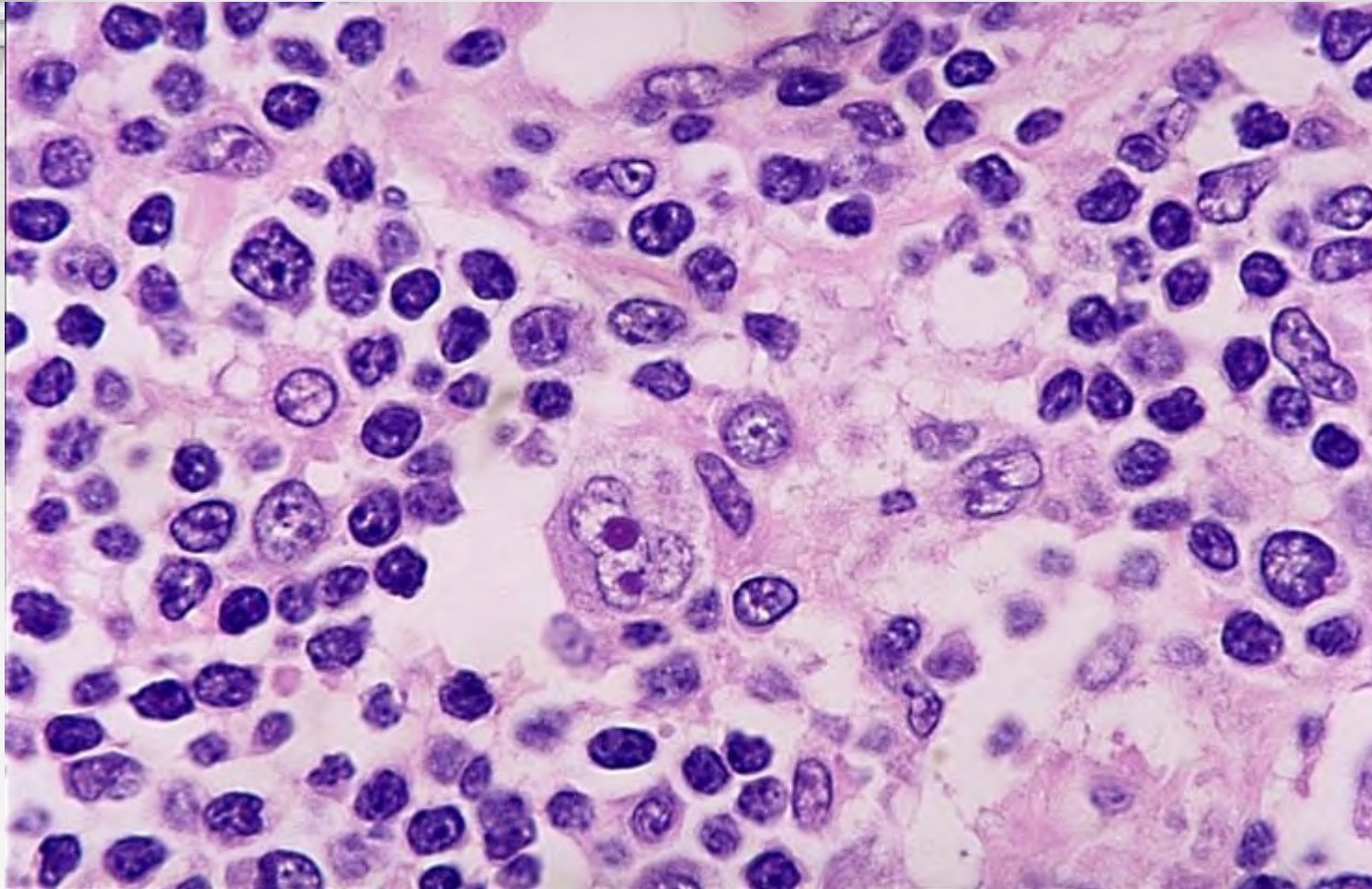
Touch preparation: binucleated large R-S cells.  
May-Giemsa staining, x 200.

## HODGKIN LYMPHOMA



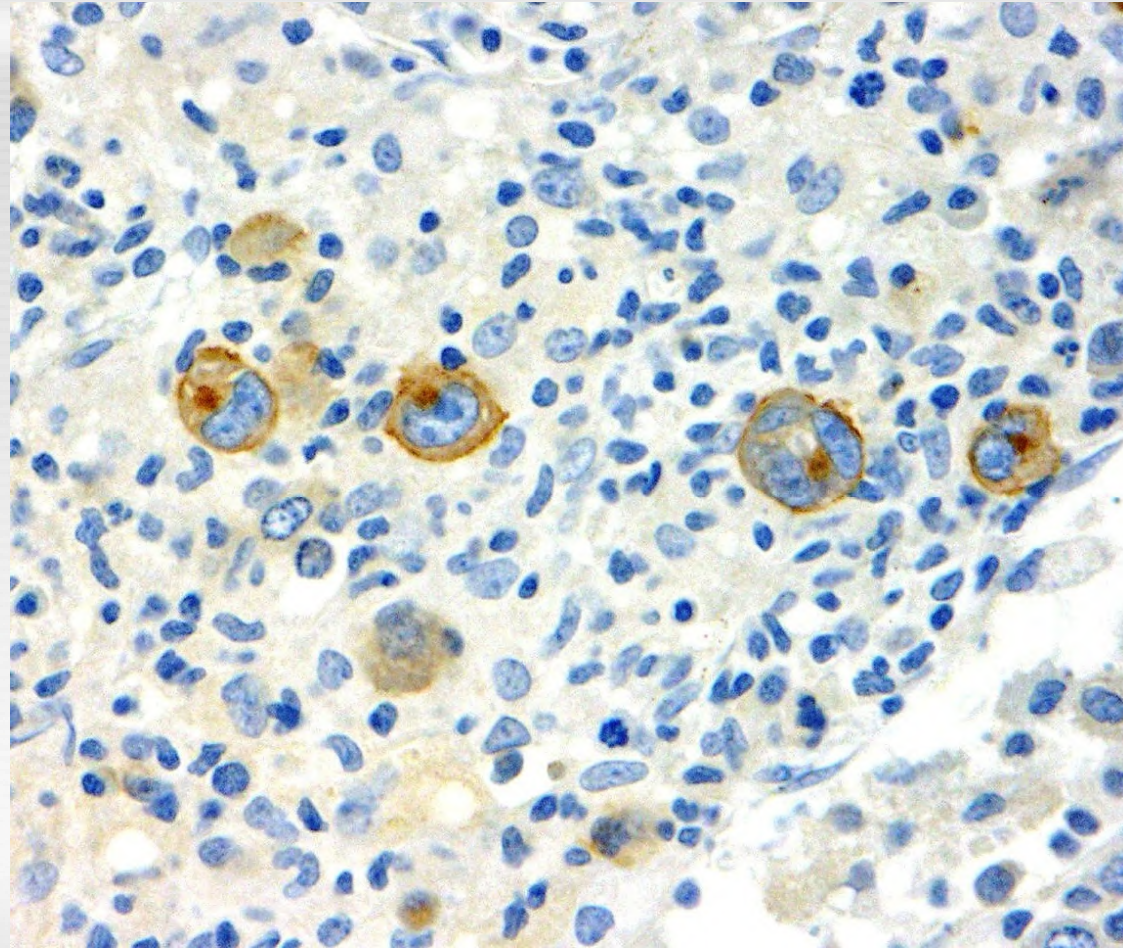
Touch preparation: binucleated large R-S cells.  
May-Giemsa staining, x 1000.

## *HODGKIN LYMPHOMA*



Lymph node.  
Hematoxylin and eosin stain, x1000

## HODGKIN LYMPHOMA



Immune phenotyping: large R-S cells positive for CD30.  
May-Giemsa staining, x 1000.

## FIRST-LINE CHEMOTHERAPY REGIMENS FOR HODGKIN LYMPHOMA

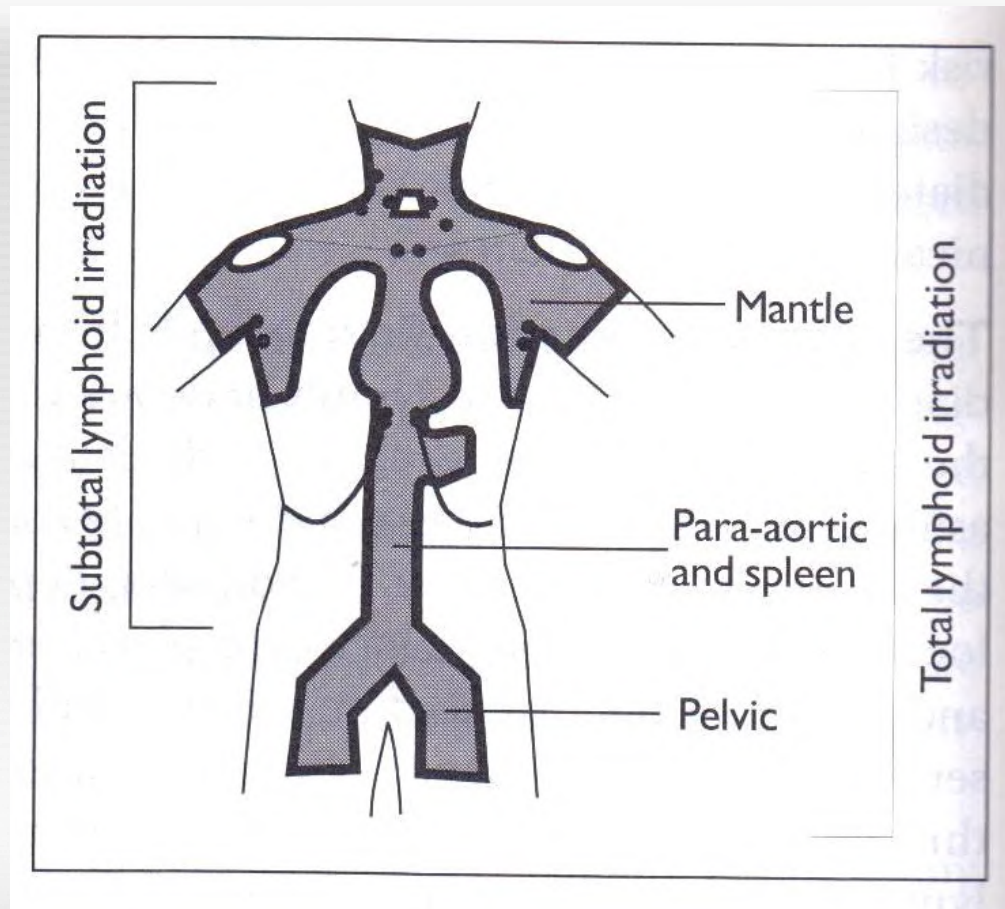
(Byrne B.J., Gockerman J.P. The Oncologist 2007;12:156–167)

Regimen/reference	Chemotherapy
ABVD [15]	25 mg/m <sup>2</sup> doxorubicin; days 1, 15 10 U/m <sup>2</sup> bleomycin; days 1, 15 6 mg/m <sup>2</sup> vinblastine; days 1, 15 375 mg/m <sup>2</sup> dacarbazine; days 1, 15
BEACOPP [13]	650 mg/m <sup>2</sup> cyclophosphamide; day 1 25 mg/m <sup>2</sup> doxorubicin; day 1 100 mg/m <sup>2</sup> etoposide; days 1–3 100 mg/m <sup>2</sup> procarbazine; days 1–7 40 mg/m <sup>2</sup> prednisone; days 1–14 1.4 mg/m <sup>2</sup> vincristine (2-mg maximum); day 8 10 U/m <sup>2</sup> bleomycin; day 8
Escalated BEACOPP [13]	1200 mg/m <sup>2</sup> cyclophosphamide; day 1 35 mg/m <sup>2</sup> doxorubicin; day 1 200 mg/m <sup>2</sup> etoposide; days 1–3 100 mg/m <sup>2</sup> procarbazine; days 1–7 40 mg/m <sup>2</sup> prednisone; days 1–14 1.4 mg/m <sup>2</sup> vincristine (2-mg maximum); day 8 10 U/m <sup>2</sup> bleomycin; day 8 G-CSF <sup>†</sup> from day 8
MOPP [16]	6 mg/m <sup>2</sup> mechlorethamine; days 1, 8 1.4 mg/m <sup>2</sup> vincristine; days 1, 8 100 mg/m <sup>2</sup> procarbazine daily; days 1–14 40 mg/m <sup>2</sup> prednisone; days 1–14
Stanford V [10]	<b>Weeks 1, 3, 5, 7, 9, and 11</b> 25 mg/m <sup>2</sup> doxorubicin 6 mg/m <sup>2</sup> vinblastine 6 mg/m <sup>2</sup> mechlorethamine; weeks 1, 5, and 9 only <b>Weeks 2, 4, 6, 8, 10, and 12</b> 1.4 mg/m <sup>2</sup> vincristine (2-mg maximum) 5 U/m <sup>2</sup> bleomycin <b>Weeks 3, 7, and 11 for 2 consecutive days</b> 60 mg/m <sup>2</sup> etoposide <b>Weeks 1–12</b> 40 mg/m <sup>2</sup> prednisone every other day for 10 weeks and then tapered by 10 mg every other day; followed by consolidative irradiation

Abbreviation: G-CSF, granulocyte cell-stimulating factor.

## ***EXTENDED RADIATION FIELDS USED FOR TREATMENT OF CLASSIC HODGKIN LYMPHOMA***

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach. 8th Edition.  
New York: CMP Healthcare Media)





## **NON-HODGKIN'S LYMPHOMAS**

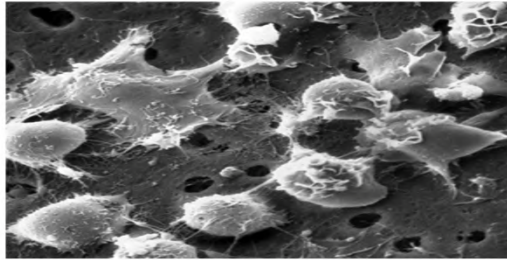
**NON-HODGKIN'S LYMPHOMAS (HL)** are a heterogenous group of neoplastic disorders originating from the extramedullary hematopoietic lymphoid cells.

The incidence of NHL in Republic of Moldova constitutes 4.1 per 100 000 population. Age-adjusted incidence rate are somewhat higher in males (4.7) than in females (3.6). Age-specific incidence rates show a logarithmic rise with increasing age. The mean age at diagnosis is 45 to 55 years. Considerable geographic and racial variations in overall incidence as well as histologic and immunologic subtypes are found worldwide, with a lower incidence of follicular lymphomas in China and Japan.

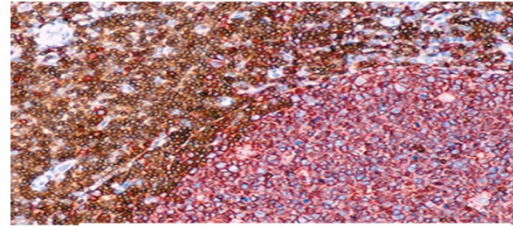
**Pathogenesis.** The heterogeneity of NHL suggests that a variety of factors including genetic abnormalities, immune disturbances, infectious agents, and other events interact in their pathogenesis. While morphology correlates with immunologic cell type (immunophenotyping), cells of identical appearance may be of types as disparate as T and B cells. Each of histologic types of NHL has its own prototype of normal cell, which NHL develop from after malignant transformation.

The development of NHL is unifocal. The morphological composition of tumour determines the clinical course of NHL. Under certain conditions, low-grade histologic types can turn into high-grade histologic types and nodular forms of tumour growth – in those diffuse. In spite of the tumour progression, a number of neoplastic cells may retain initial caryotypical and immunologic markers.

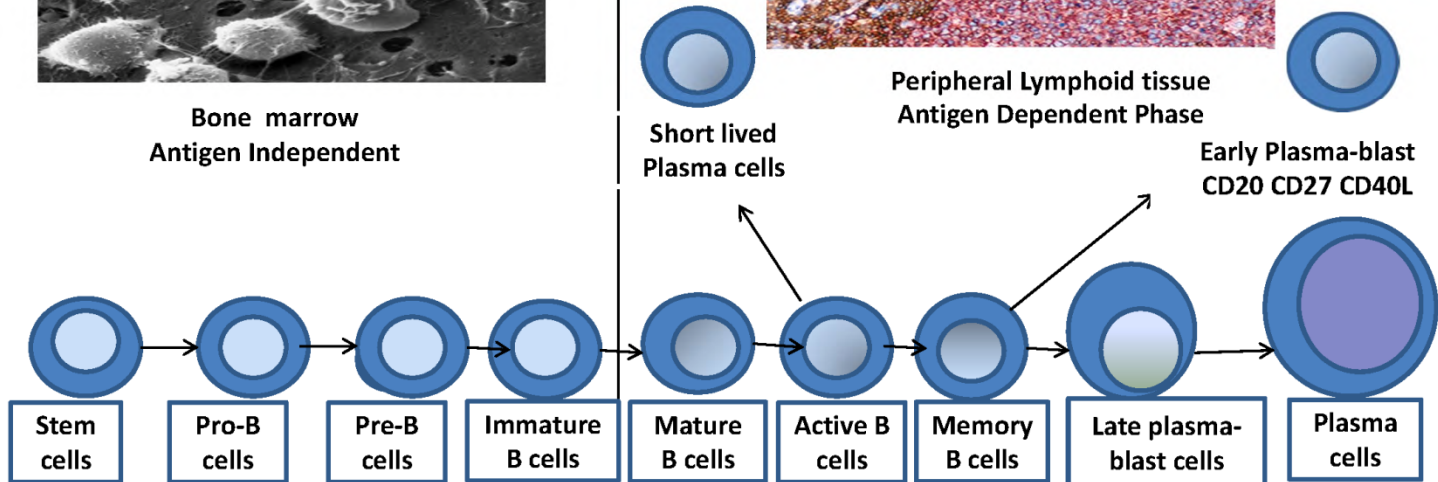
# MULTI-STEP TRANSFORMATION OF LYMPHOID CELLS



Bone marrow  
Antigen Independent



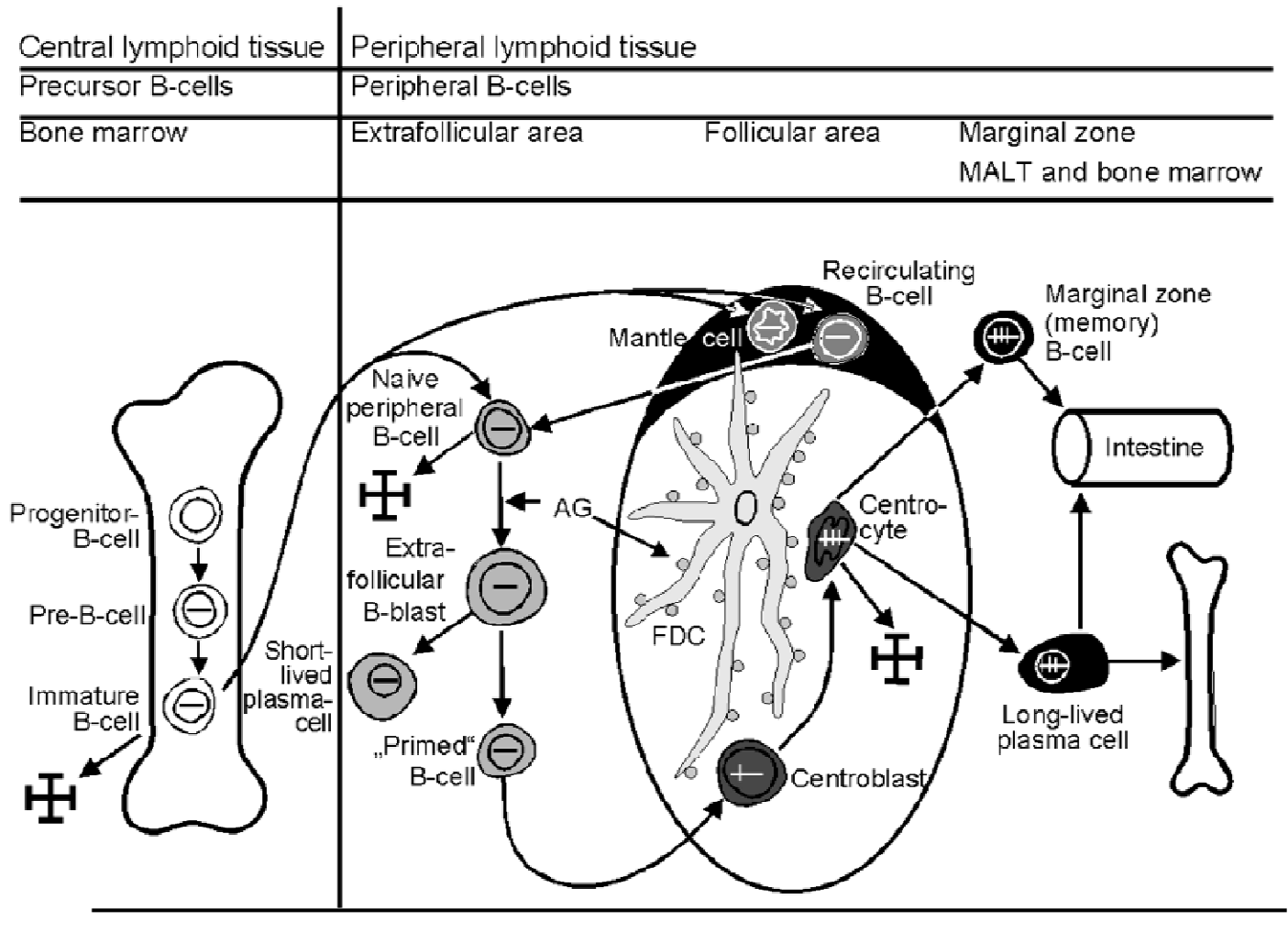
Peripheral Lymphoid tissue  
Antigen Dependent Phase



Cell Surface Antigens	
CD19	←—————→
CD20	←—————→ •
CD27	←—————→ •
Serum BAFF and CD38	←—————→ •
CD138	←—————→ •

## EVENTS IN B-CELL DEVELOPMENT

(American Society of Hematology, New Approaches to Lymphoma Diagnosis. Hematology 2001)



## EVENTS IN B-CELL DEVELOPMENT

(American Society of Hematology, New Approaches to Lymphoma Diagnosis. Hematology 2001)

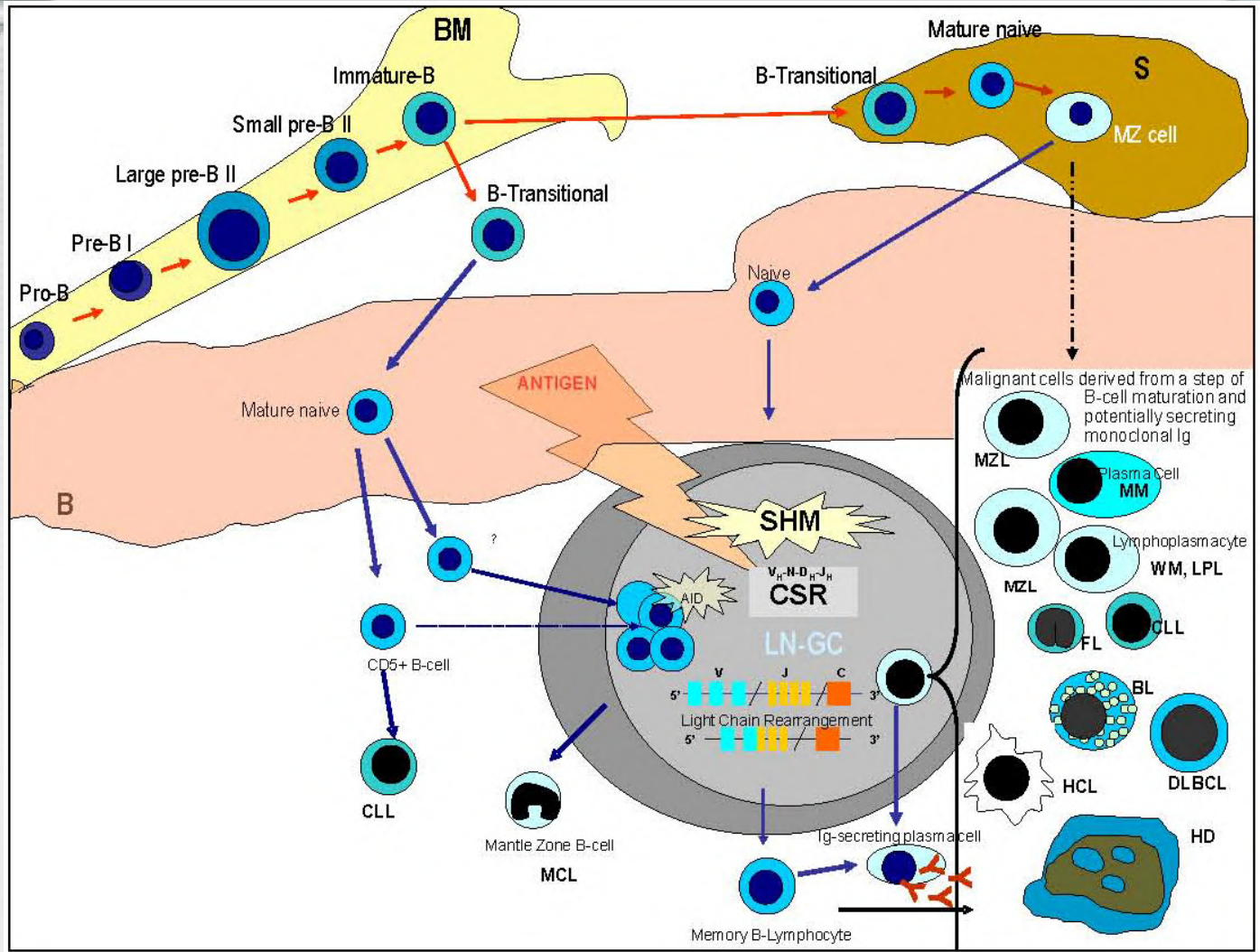
The development and maturation process of B cells begins in the bone marrow. Here, the “pre-B cell” arises from the “progenitor (Pro) B cell” following rearrangement of the immunoglobulin heavy chain gene (symbolized with horizontal lines in black). Subsequently, rearrangement of the light chain genes occurs resulting in the expression of the whole immunoglobulin molecule on the cell surface, serving as an antigen receptor. With the production of this “immature” B cell, the initial phase of B-cell development is, thereby, completed. The “immature” B cell is so defined since it is unable to initiate an immune response following the presentation of a foreign antigen. The B-cell attains this ability only on leaving the bone marrow, passing through the blood stream and entering the peripheral lymphoid tissue. Here, the B cell migrates to the outer region of the lymph node in the “primary” follicles and, later, to the follicle mantles. This differentiation step is associated with the additional expression of IgD. These IgM+/IgD+ B cells are known as “naive mature B-cells”. When these cells come into contact with antigen (AG), which can bind to their immunoglobulin molecules, they transform into proliferating extrafollicular B blasts, from which short-lived plasma cells and “antigen-induced” or “primed” B cells are derived. These “primed” B cells initiate and maintain the germinal center reaction, during which they transform into rapidly proliferating centroblasts. During the mitotic proliferation and differentiation of the centroblasts into centrocytes, somatic mutations in the variable region of the immunoglobulin genes are inserted in a randomized manner (the mutations are represented by vertical lines). The centrocytes with advantageous mutations (i.e. those which lead to an increase in the affinity of the immunoglobulin receptor) differentiate further, passing out of the germinal centre into long-lived plasma cells or into “memory” B cells. The latter remain in the marginal zone. FDC, follicular dendritic cell;  $\mp$ , apoptosis

As a result of the differentiation phases of B-cells and of the somatic mutation process, 3 major different mature forms of B-cells can be identified:

- Naive mature B-cells (recirculating and sessile subtypes)
- Germinal center B-cells (centroblasts and centrocytes)
- Post germinal center B-cells which include memory B cells and long-lived plasma cells

From all of these different B-cell forms, malignant B-cell lymphomas arise, which distinguish themselves clinically and which are characterized in their biological behavior not only by the transformation event but also by the inherent characteristics of the cell of origin. Classical Hodgkin lymphomas, in which the phenotypical and clinical features are predominantly determined by the transformation event, are an exception to this rule.

# MULTI-STEP TRANSFORMATION OF LYMPHOID CELLS



## LOCALIZATIONS OF PRIMARY TUMOR FOCUS IN NON-HODGKIN LYMPHOMAS

<i>Localizarea primară</i>	<i>Numărul de bolnavi</i>	
	<i>Abs.</i>	<i>%</i>
<b>Ganglionii limfatici</b>	<b>260</b>	<b>51,0</b>
periferici	173	33,9
mediastinali	12	2,4
retroperitoneali și abdominali	39	7,6
grupa neidentificată	36	7,1
<b>Localizări extranodale</b>	<b>214</b>	<b>42,0</b>
inelul Waldayer	80	15,7
tractul gastrointestinal	68	13,3
splina	21	4,1
oasele, pielea, țesuturile moi	23	4,5
plămâni, pleura	2	0,4
glanda mamară	4	0,8
alte localizări	16	3,2
nu a fost identificată	36	7,0
<b>Total</b>	<b>510</b>	<b>100,0</b>

*PRIMARY CUTANEOUS NON-HODGKIN LYMPHOMA*



*PRIMARY EYELID NON-HODGKIN LYMPHOMA*

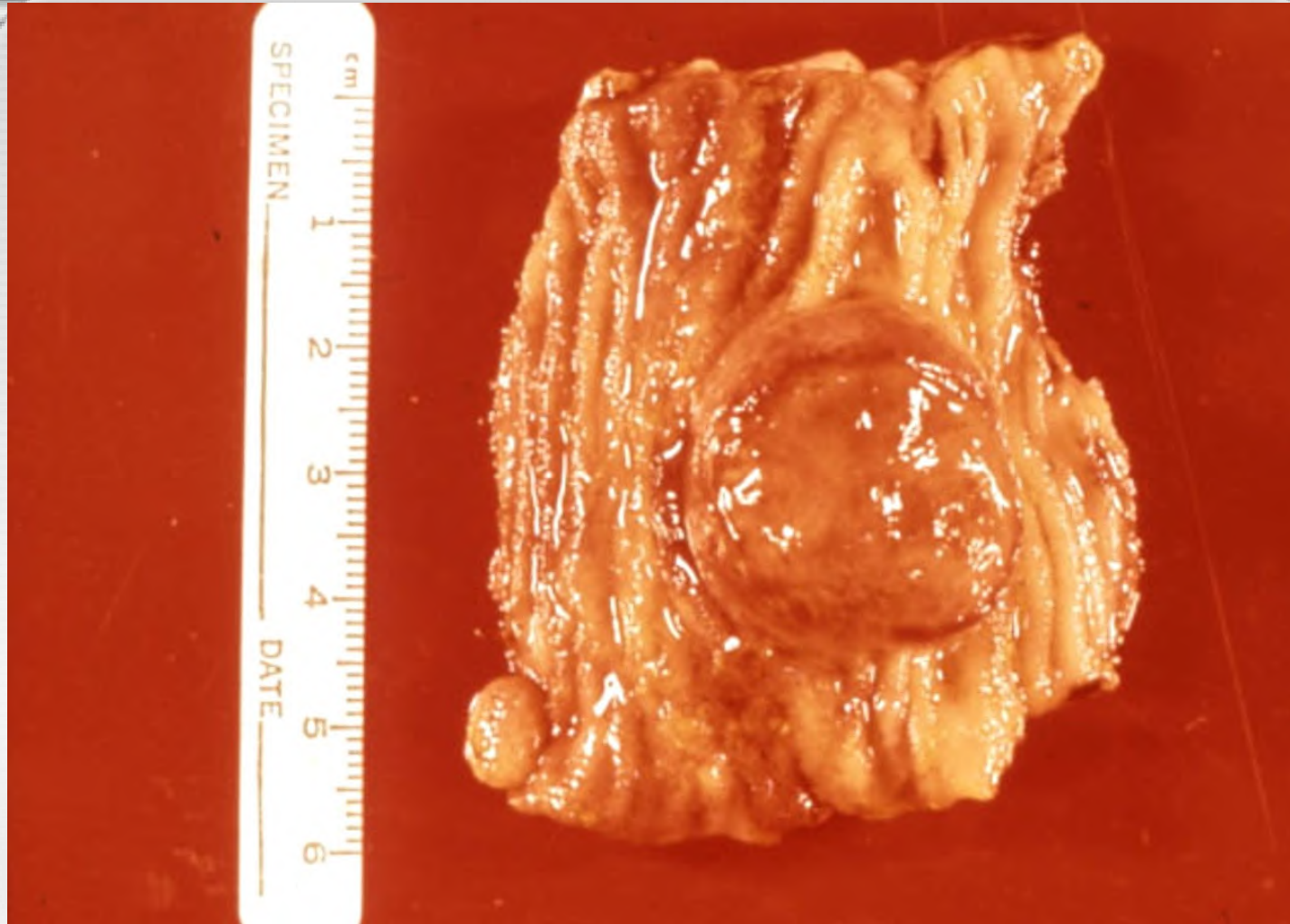




*PRIMARY EYELID NON-HODGKIN LYMPHOMA*

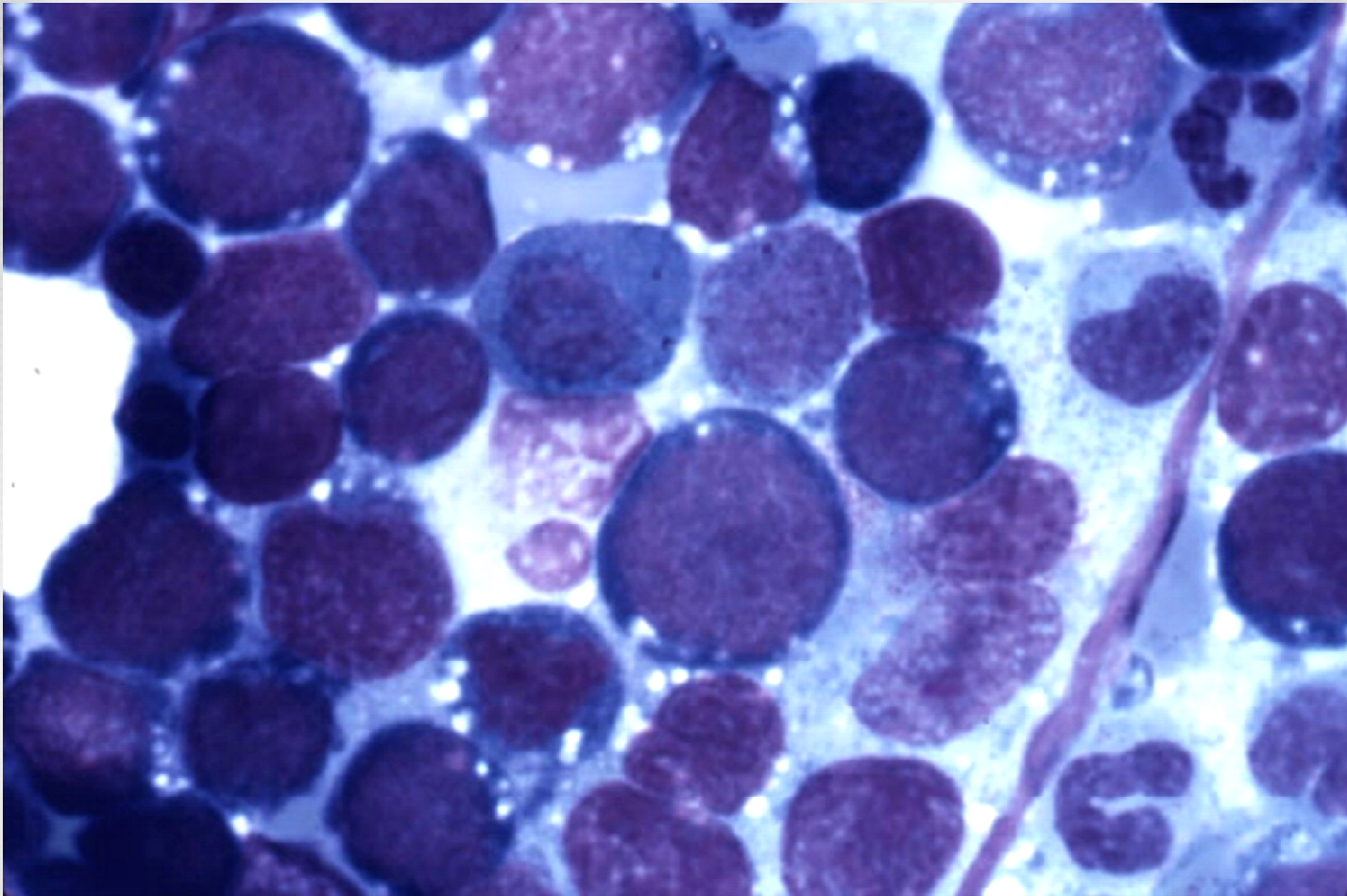


*NON-HODGKIN LYMPHOMA, BURKITT TYPE*



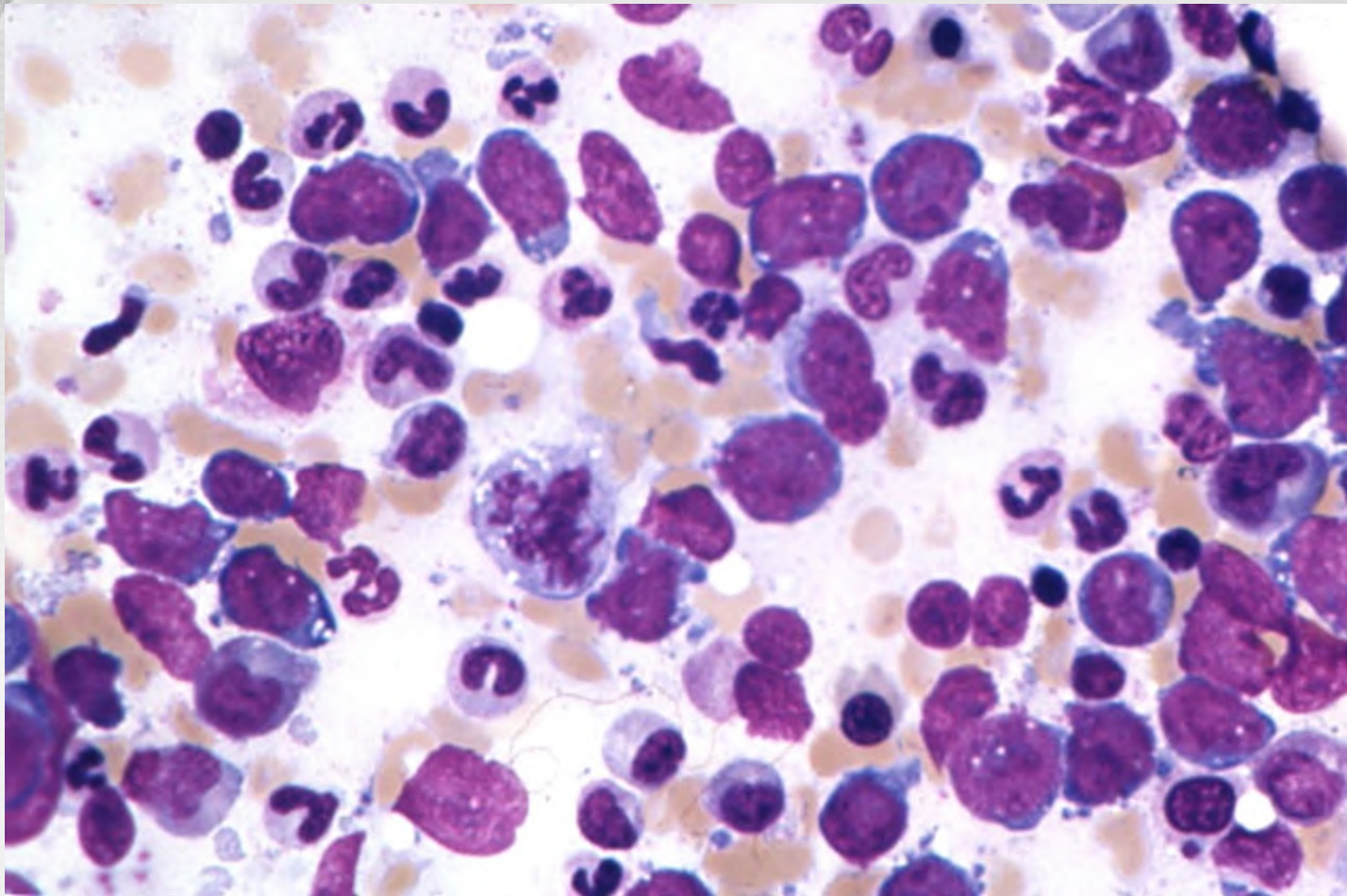
**Obstruction lesion in ileum, a common presenting clinical feature in sporadic Burkitt lymphoma**

*NON-HODGKIN LYMPHOMA, BURKITT TYPE*



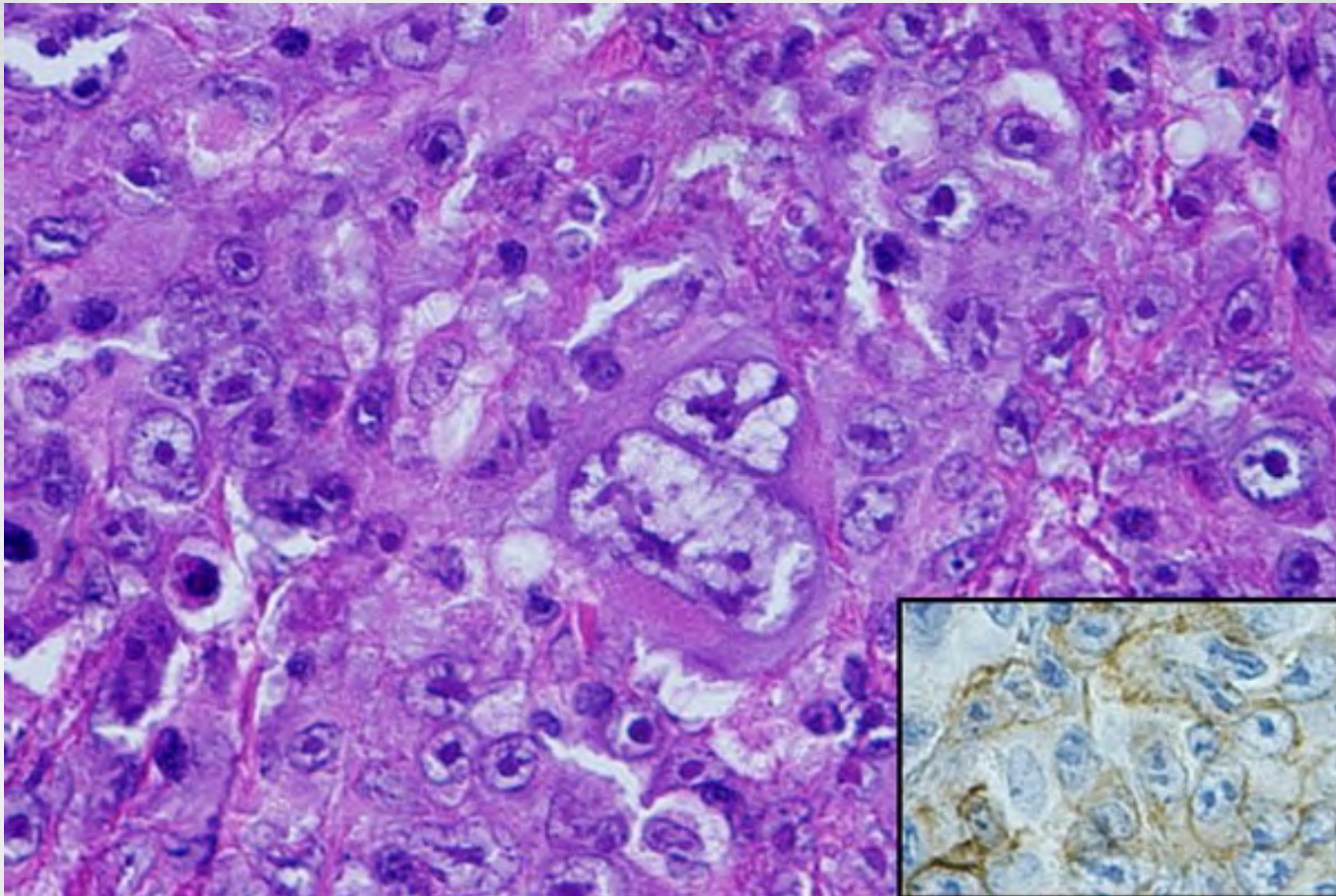
Cytology showing deeply basophilic vacuolated cytoplasm - vacuoles contain lipid.  
May-Giemsa staining, x 1000.

*NON-HODGKIN LYMPHOMA, BURKITT TYPE*



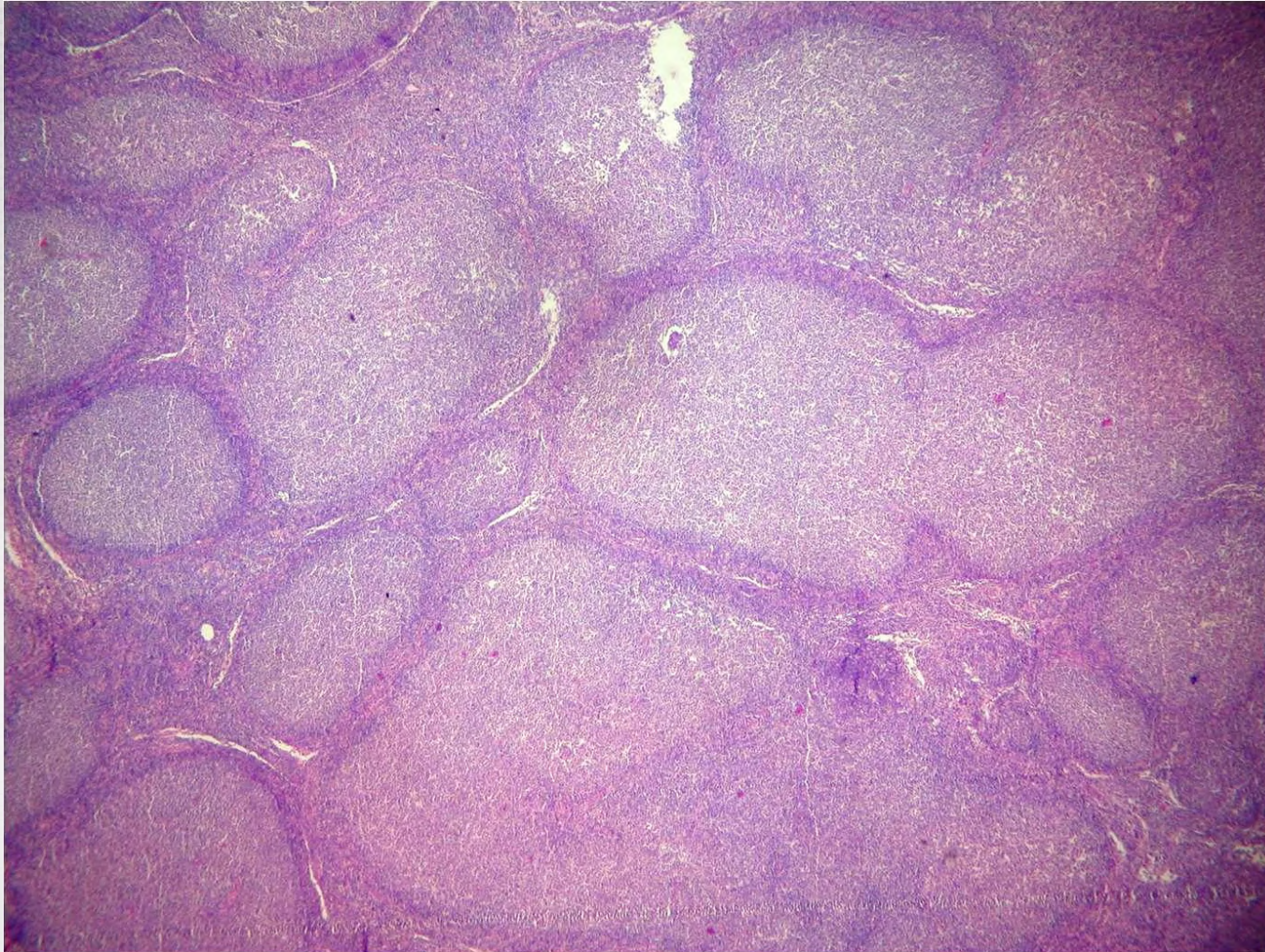
Bone marrow aspirate with admixture of Burkitt lymphoma cells.  
May-Giemsa staining, x 1000.

*DIFFUSE LARGE B-CELL NON-HODGKIN LYMPHOMA, ANAPLASTIC TYPE*



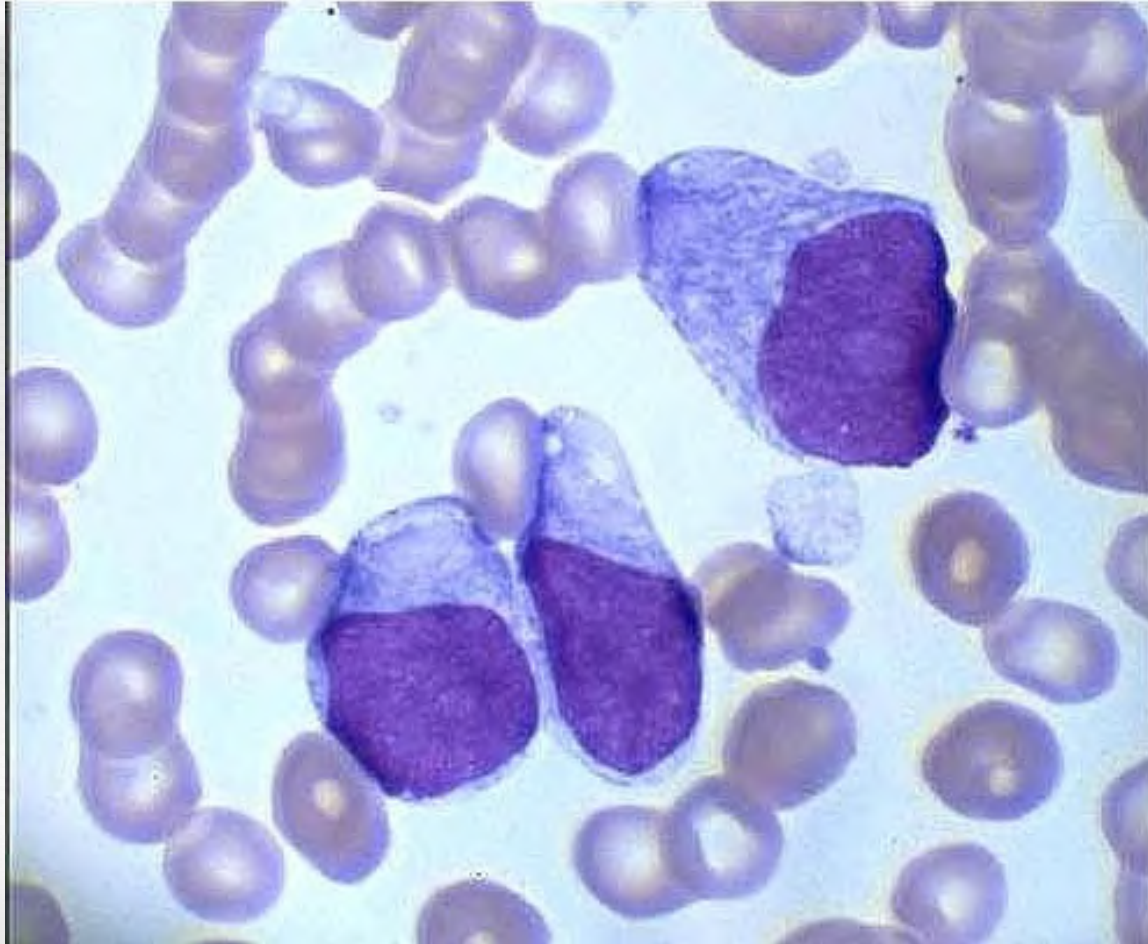
Lymph node biopsy.  
Hematoxylin and eosin stain, x1000.

*FOLLICULAR NON-HODGKIN LYMPHOMA*



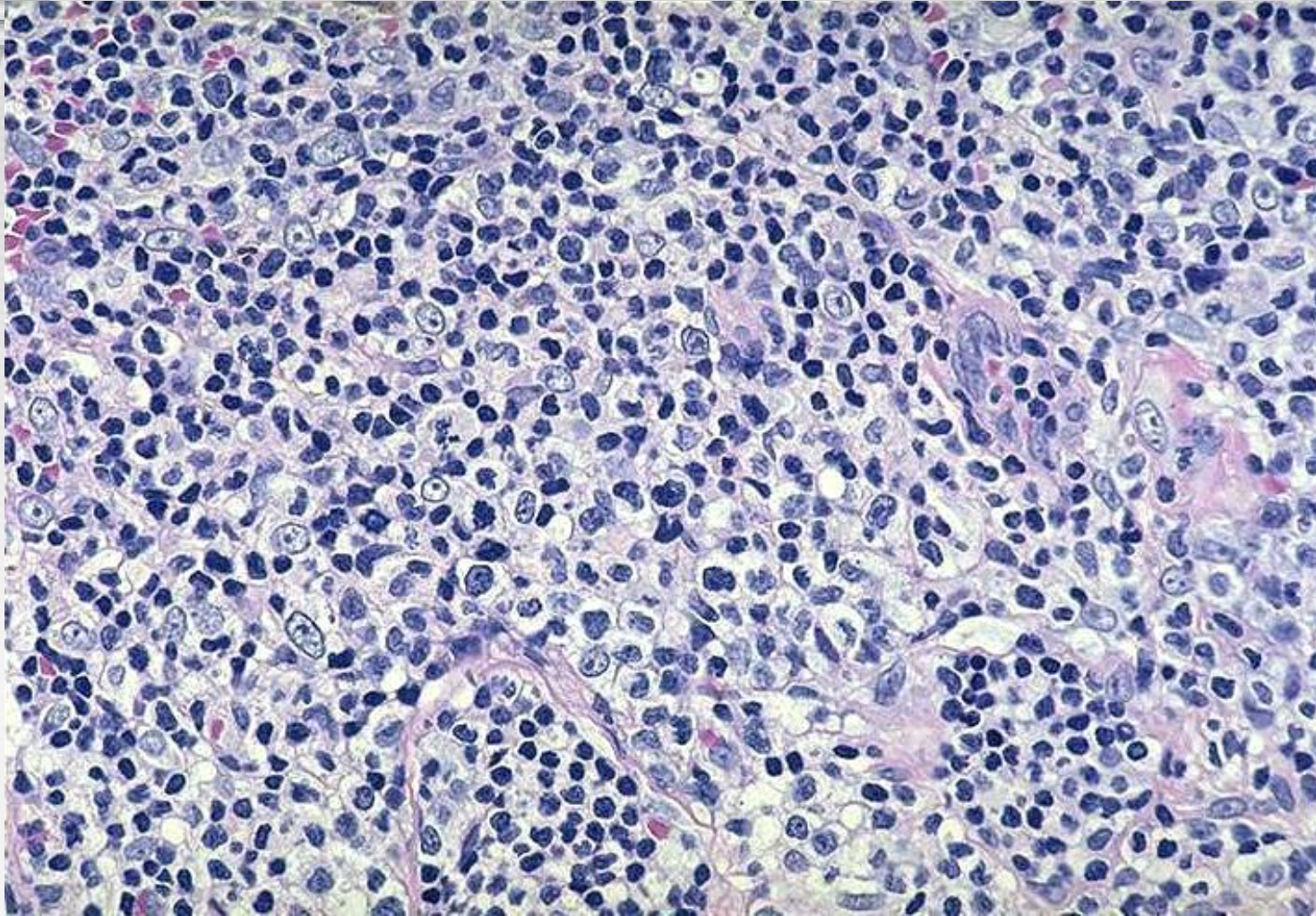
May-Giemsa staining, x 100

***BONE MARROW SMEAR IN NON-HODGKIN LYMPHOMA***



May-Giemsa staining, x 1000

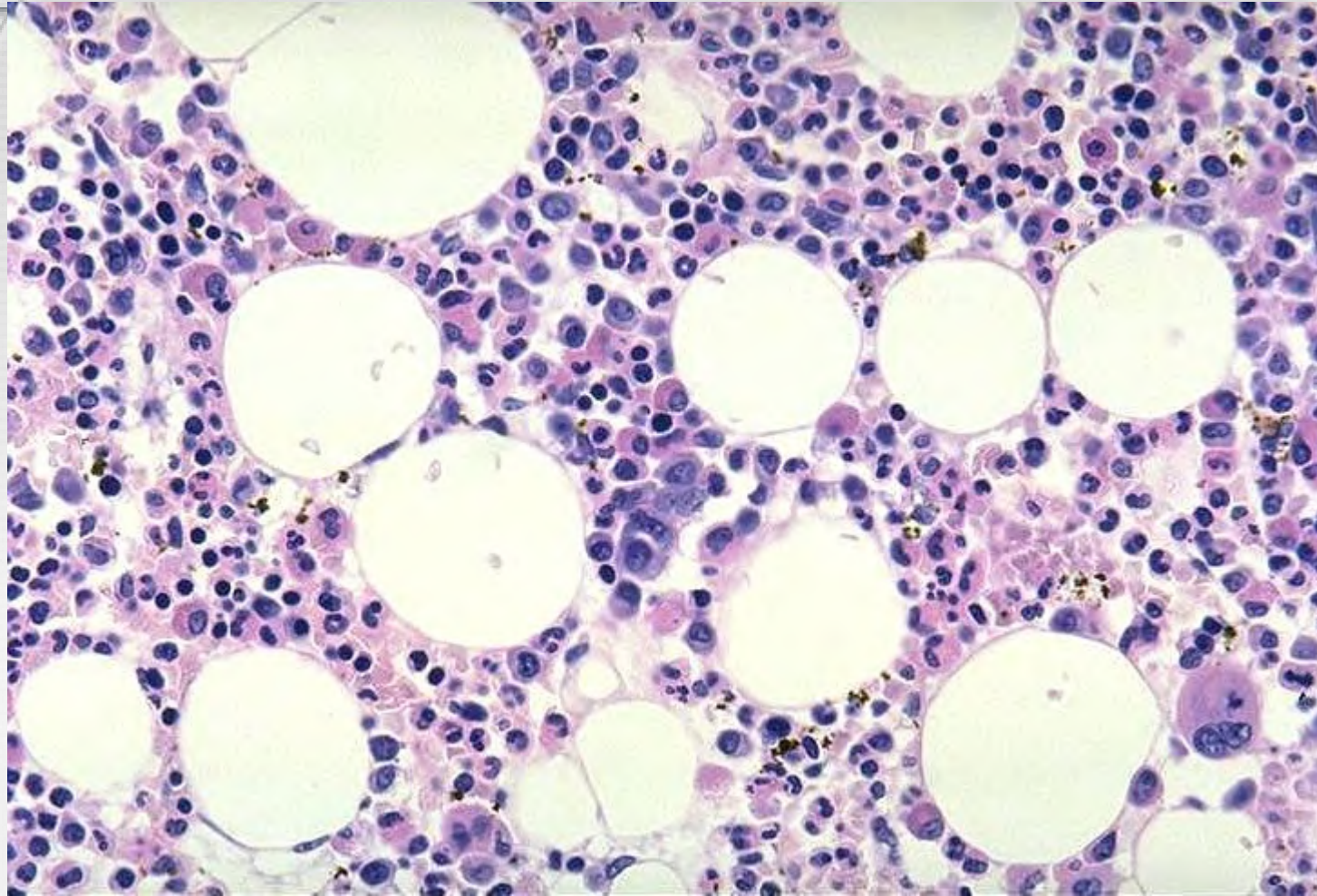
*CUTANEOUS T-CELL NON-HODGKIN LYMPHOMA*



Hematoxylin and eosin stain, x400

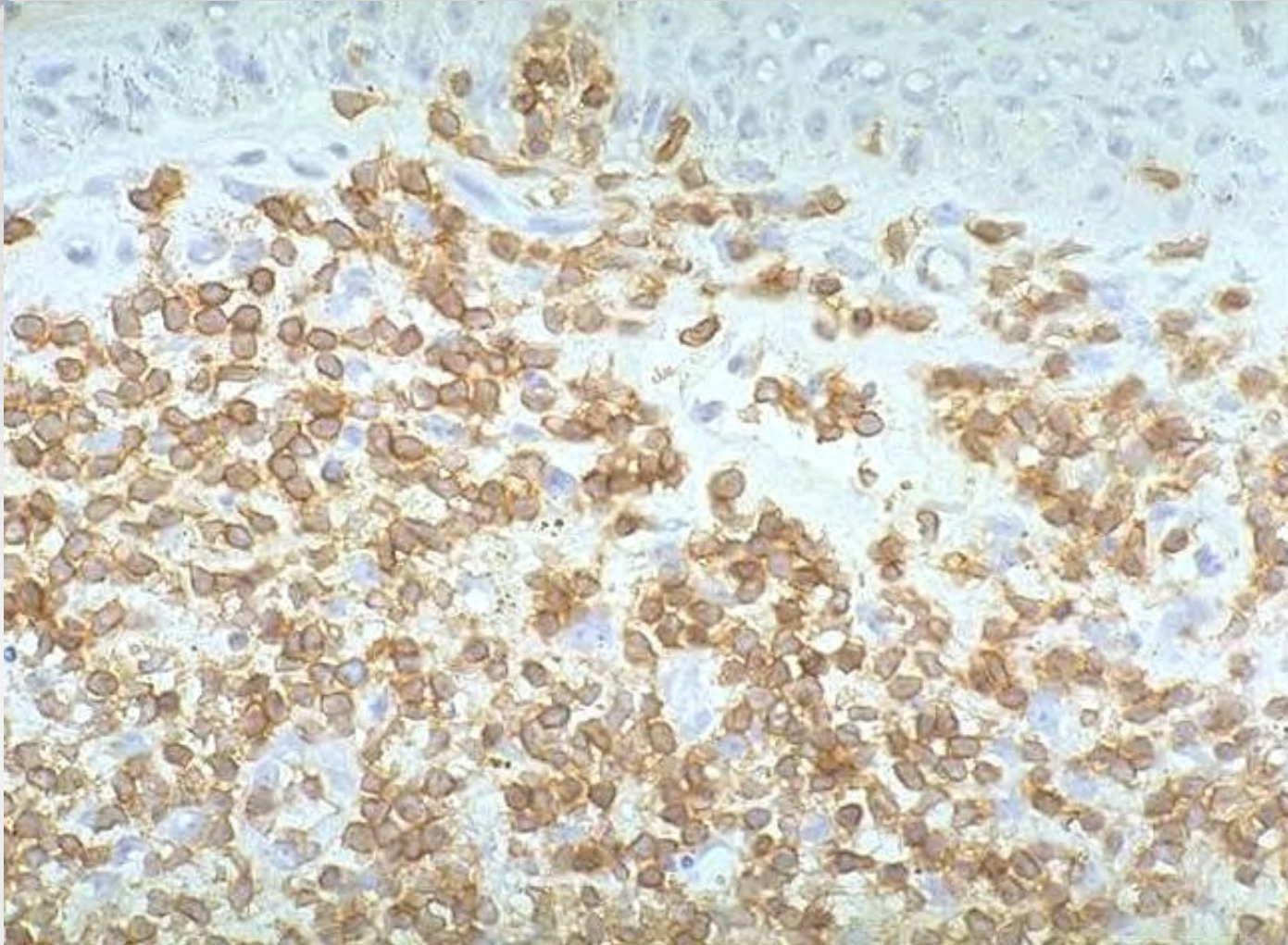


*CUTANEOUS T-CELL NON-HODGKIN LYMPHOMA*



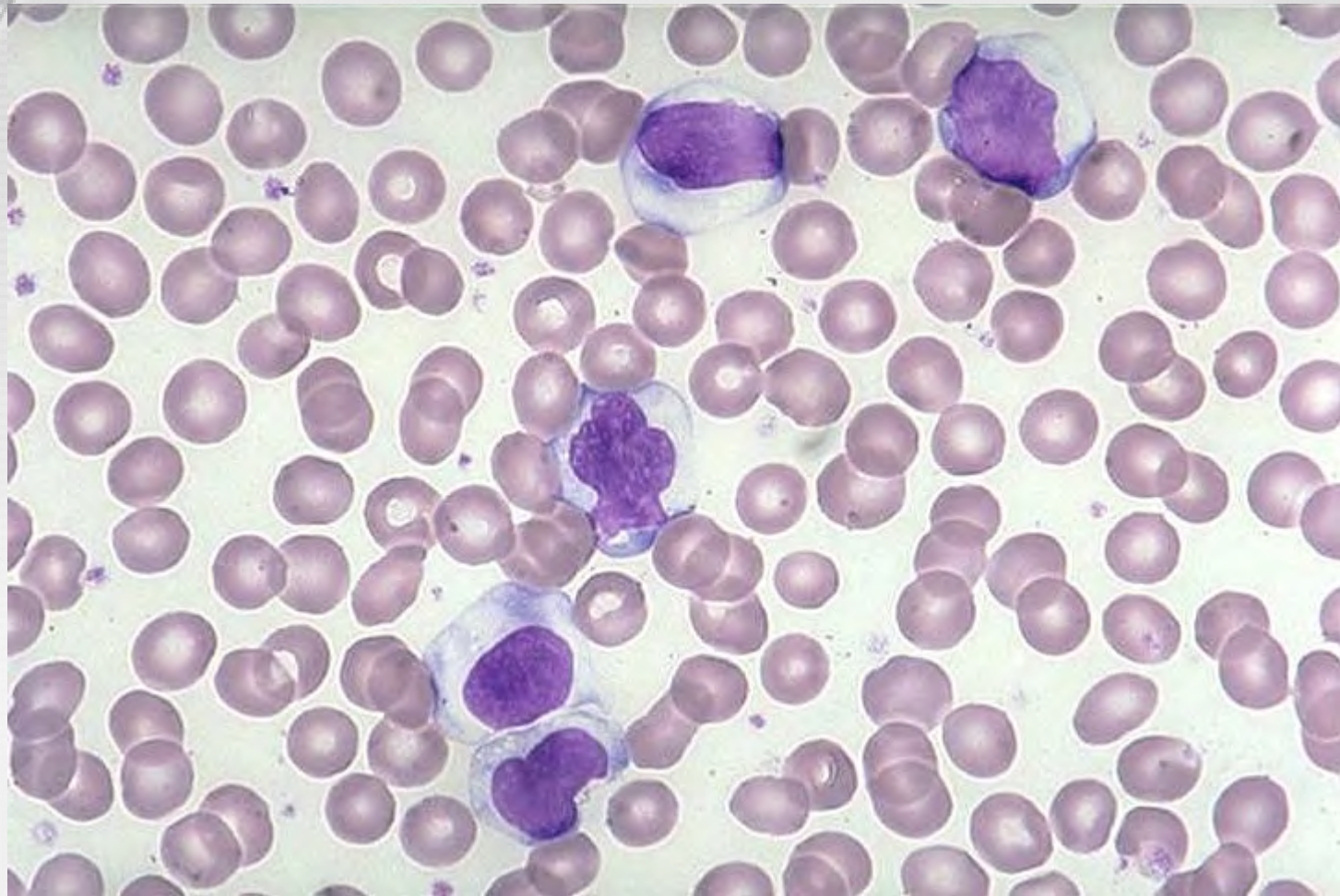
Clot section.  
Hematoxylin and eosin stain, x400.

*CUTANEOUS T-CELL NON-HODGKIN LYMPHOMA*



Immunostain\_antibody CD3(Leu4), x400

*CUTANEOUS T-CELL NON-HODGKIN LYMPHOMA*



Blood smear.  
May-Giemsa stain, x1000.

# IMMUNOPHENOTYPIC AND GENETIC FEATURES OF COMMON B-CELL NEOPLASMS

(American Society of Hematology, New Approaches to Lymphoma Diagnosis. Hematology 2001)

Neoplasm	Slg; clg	CD5	CD10	Bcl6	CD23	CD43	CD103	Cyclin D1	CD 138	Genetic Abnormality	Immuno-globulin Genes*
B-SLL/CLL	+; -/+	+	-	-	+	+	-	-	-	trisomy 12; 13q	R,U (50%); M (50%)
Lymphoplasmacytic lymphoma	++;	-	-	-	-	+/-	-	-	-/+	t(9;14); del 6(q23)	R,M
Hairy cell leukemia	+;-	-	-	-	-	+	++	+/-	-	none known	R,M
Plasma cell myeloma	-;+	-	-/+	-	-	-/+	-	-/+	+	t(4:14), t(6:14) t(14;16), t(1;14)	R,M
Splenic marginal zone lymphoma	+; -/+	-	-	-	-	-	+	-	-	none known	R,M
Follicular lymphoma	+;-	-	+/-	+	-/+	-	-	-	-	t(14;18); bcl-2	R,M,O
Mantle cell lymphoma	+;-	+	-	-	-	+	-	+	-	t(11;14); bcl-1	R,U
MALT lymphoma	+; +/-	-	-	-	-/+	-/+	-	-	-	+3, t(11;18); API2/MLT1	R,M,O
Diffuse large B-cell lymphoma	+/-; -/+	-	-/+	+/-	NA	-/+	NA	-	-/+	t(14;18);t(8;14) 3q27; BCL2, cMYC, BCL6	R,M
Burkitt lymphoma	+;-	-	+	+	-	-	NA	-	-	t(8;14), t(2;8), t(8;22); cMYC; EBV-/+	R,M

Abbreviations: R, rearranged; M, mutated; NK, natural killer cell; U, unmutated; O, ongoing mutations; NA, not available

Key: + = >90% positive; +/- = > 50% positive; -/+ = < 50% positive; - = < 10% positive

## IMMUNOPHENOTYPIC AND GENETIC FEATURES OF COMMON T-CELL NEOPLASMS

(American Society of Hematology, New Approaches to Lymphoma Diagnosis. Hematology 2001)

Neoplasm	CD3 (S;C)	CD5	CD7	CD4	CD8	CD30	TCR	NK <sup>16, 56</sup>	Cytotoxic granule
T-prolymphocytic leukemia	+	-	+,+	+/-	-/+	-	$\alpha\beta$	-	-
T-large granular lymphoproliferative disease	+	-	+,+	-	+	-	$\alpha\beta$	+,-	+
NK large granular lymphoproliferative disease	-	-	+, -	-	+/-	-	-	-,+	+
Extranodal NK/T-cell lymphoma	-;+	-	-/+	-	-	-	-	NA,+	+
Hepatosplenic T-cell lymphoma	+	-	+	-	-	-	$\gamma\delta \gg \alpha\beta$	+,-/+	+
Enteropathy-type T-cell lymphoma	+	+	+	-	+/-	+/-	$\alpha\beta \gg \gamma\delta$	-	+
Mycosis fungoides	+	+	-/+	+	-	-	$\alpha\beta$	-	-
Cutaneous anaplastic large cell lymphoma	+	+/-	+/-	+/-	-	++	$\alpha\beta$	-	-/+
Subcutaneous panniculitis-like T-cell	+	+	+	-	+	-/+	$\alpha\beta > \gamma\delta$	-, +/-	+
Peripheral T-cell lymphoma, unspecified	+/-	+/-	+/-	+/-	-/+	-/+	$\alpha\beta > \gamma\delta$	-/+	-/+
Angioimmunoblastic	+	+	+	+/-	-/+	-	$\alpha\beta$	-	NA
Primary systemic anaplastic large cell lymphoma	+/-	+/-	NA	-/+	-/+	++	$\alpha\beta$	-	+

Abbreviations: R, rearranged; M, mutated; NK, natural killer cell; U, unmutated; O, ongoing mutations; TCR, T-cell receptor gene; Ig, immunoglobulin; NA, not available

Key: + = >90% positive; +/- = > 50% positive; -/+ = < 50% positive; - = < 10% positive; Cytotoxic granule = TIA-1, perforin, and/or granzyme

\* Mutations in the Ig gene V region indicate exposure to antigen.

# INTERNATIONAL PROGNOSTIC INDEX AND AGE-ADJUSTED INDEX FOR AGGRESSIVE LYMPHOMA PATIENTS TREATED WITH DOXORUBICIN-CONTAINING COMBINATION CHEMOTHERAPY

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach. 8th Edition. New York: CMP Healthcare Media)

## IPI: Risk factors:

- Age >60 years
- LDH > normal
- ECOG performance status 2–4
- Stage III or IV
- Two or more extranodal sites of disease

Risk group	Risk factors (Sum)	CR (%)	5-year OS (%)
Low	0–1	87	73
Low-intermediate	2	67	51
High-intermediate	3	55	43
High	4–5	44	26

## Age-adjusted IPI for age <60 years:

### Risk factors:

- LDH > normal
- Performance status 2–4
- Stage III–IV

Low	0	92	83
Low-intermediate	1	78	69
High-intermediate	2	57	46
High	3	46	32

From Shipp. Blood 1994;83:1165–73.

CR = complete response; ECOG = Eastern Cooperative Oncology Group; OS = overall survival.

## **CHEMOTHERAPY REGIMENS, COMMONLY USED IN NON-HODGKIN LYMPHOMAS**

(Williams M.E., Kahn M.J., American Society of Hematology Self-Assessment Program. Blackwell Publishing: 2005)

Newly diagnosed patients	Relapsed and refractory patients
<b>CVP:</b>	<b>ICE:</b>
Cyclophosphamide	Ifosfamide
Vincristine	Carboplatin
Prednisone	Etoposide
<b>CHOP:</b>	<b>DHAP:</b>
CVP plus doxorubicin	Dexamethasone
<b>R-CHOP:</b>	High-dose araC
CHOP plus rituximab	Cis-platinum
<b>MACOP-B:</b>	<b>ESHAP:</b>
Methotrexate/leucovorin	Etoposide
Doxorubicin	Methylprednisolone
Cyclophosphamide	High-dose araC
Vincristine	Cis-platinum
Prednisone	<b>EPOCH:</b>
Bleomycin	Etoposide*
<b>M-BACOD:</b>	Vincristine*
Methotrexate/leucovorin	Doxorubicin*
Bleomycin	Cyclophosphamide
Doxorubicin	Prednisone
Cyclophosphamide	
Vincristine	
Dexamethasone	
<b>ProMACE-CytaBOM:</b>	
Prednisone	
Doxorubicin	
Cyclophosphamide	
Etoposide	
Cytosine arabinoside (araC)	
Bleomycin	
Vincristine	
Methotrexate/leucovorin	

\*Continuous infusion × 4 days/cycle.

## COMBINATION CHEMOTHERAPY IN NON-HODGKIN LYMPHOMAS: REGIMENS, MODE OF ADMINISTRATION AND DOSES

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach.  
8th Edition. New York: CMP Healthcare Media)

Regimen	Dose	Route and frequency
<b>CVP ± Rituximab</b>		
Cyclophosphamide	400 mg/m <sup>2</sup>	750-1,000 mg/m <sup>2</sup> on day 1
Vincristine	1.4 mg/m <sup>2</sup>	IV on day 1 (maximum, 2 mg)
Prednisone	100 mg or 100 mg/m <sup>2</sup>	PO on days 1-5
Rituximab	375 mg/m <sup>2</sup>	IV on day 1
<i>Repeat treatment every 21 days.</i>		
<b>CHOP ± Rituximab</b>		
Cyclophosphamide	750 mg/m <sup>2</sup>	IV on day 1
Doxorubicin HCl	50 mg/m <sup>2</sup>	IV on day 1
Oncovin	1.4 mg/m <sup>2</sup>	IV on day 1 (maximum, 2 mg)
Prednisone	40 mg/m <sup>2</sup> or 100 mg/day or 100 mg/m <sup>2</sup> /day	PO on days 1-5
<i>Repeat treatment every 21 days.</i>		
<b>CHOEP ± Rituximab</b>		
Cyclophosphamide	750 mg/m <sup>2</sup>	IV on day 1
Doxorubicin	50 mg/m <sup>2</sup>	IV on day 1
Etoposide	100 mg/m <sup>2</sup>	IV on days 1-3
Oncovin	1.4 mg/mL	IV on day 1 (maximum, 2 mg)
Prednisone	100 mg	PO on days 1-5
Rituximab	375 mg/m <sup>2</sup>	IV on day 1
<i>Repeat treatment every 21 days.</i>		
<b>C-MOPP</b>		
Cyclophosphamide	650 mg/m <sup>2</sup>	IV on days 1, 8
Oncovin	1.4 mg/m <sup>2</sup>	IV on days 1, 8
Procarbazine	100 mg/m <sup>2</sup>	PO on days 1-14
Prednisone	40 mg	PO on days 1-14
<i>Repeat treatment every 28 days.</i>		
<b>MACOP-B</b>		
Methotrexate <sup>3</sup>	400 mg/m <sup>2</sup>	IV on weeks 2, 6, 10
Adriamycin	50 mg/m <sup>2</sup>	IV on weeks 1, 3, 5, 7, 9, 11
Cyclophosphamide	350 mg/m <sup>2</sup>	IV on weeks 1, 3, 5, 7, 9, 11
Oncovin	1.4 mg/m <sup>2</sup>	IV on weeks 2, 4, 6, 8, 10, 12 (maximum, 2 mg)
Prednisone	75 mg	PO daily for 12 weeks; dose tapered over the last 15 days
Bleomycin	10 U/m <sup>2</sup>	IV on weeks 4, 8, 12
Co-trimoxazole	2 tablets	PO twice daily throughout



# COMBINATION CHEMOTHERAPY IN NON-HODGKIN LYMPHOMAS: REGIMENS, MODE OF ADMINISTRATION AND DOSES

(Pazdur R., Coia L.R., Hoskins W.J. et al., Cancer Management: A Multidisciplinary Approach. 8th Edition. New York: CMP Healthcare Media)

Regimen	Dose	Route and frequency
<b>FND</b>		
Fludarabine	25 mg/m <sup>2</sup>	IV on days 1-3
Novantrone	10 mg/m <sup>2</sup>	IV on day 1
Dexamethasone	20 mg	PO/IV on days 1-5
<i>Repeat treatment every 21-28 days depending on hematologic recovery.</i>		
<b>ProMACE-CytaBOM</b>		
Cyclophosphamide <sup>a</sup>	650 mg/m <sup>2</sup>	IV on day 1
Etoposide	120 mg/m <sup>2</sup>	IV on day 1
Adriamycin	25 mg/m <sup>2</sup>	IV on day 1
Cytarabine	300 mg/m <sup>2</sup>	IV on day 8
Bleomycin	5 U/m <sup>2</sup>	IV on day 8
Oncovin	1.4 mg/m <sup>2</sup>	IV on day 8 (maximum, 2 mg)
Methotrexate	120 mg/m <sup>2</sup>	IV on day 8
Leucovorin	25 mg/m <sup>2</sup>	PO q6h for 4 doses; start 24 hours after methotrexate
Prednisone	60 mg	PO on days 1-14
Co-trimoxazole	2 tablets	PO twice daily throughout
<i>Repeat treatment every 28 days.</i>		
<b>ProMACE-MOPP</b>		
Cyclophosphamide	650 mg/m <sup>2</sup>	IV on day 1
Etoposide	120 mg/m <sup>2</sup>	IV on day 1
Adriamycin	25 mg/m <sup>2</sup>	IV on day 1
Procarbazine	100 mg/m <sup>2</sup>	PO on days 8-14
Mechlorethamine	6 mg/m <sup>2</sup>	IV on day 8
Oncovin	1.4 mg/m <sup>2</sup>	IV on day 8 (maximum, 2 mg)
Prednisone	60 mg	PO on days 1-14
Methotrexate	500 mg/m <sup>2</sup>	IV on day 15
Leucovorin	50 mg/m <sup>2</sup>	PO q6h for 5 doses; start 24 hours after methotrexate
<i>Repeat treatment every 28 days.</i>		
<b>m-BACOD</b>		
Methotrexate	200 mg/m <sup>2</sup>	IV on days 8, 15
Leucovorin	10 mg/m <sup>2</sup>	PO q6h for 8 doses; start 24 hours after methotrexate
Bleomycin	4 U/m <sup>2</sup>	IV on day 1
Adriamycin	45 mg/m <sup>2</sup>	IV on day 1
Cyclophosphamide	600 mg/m <sup>2</sup>	IV on day 1
Oncovin	1 mg/m <sup>2</sup>	IV on day 1 (maximum, 2 mg)
Dexamethasone	6 mg/m <sup>2</sup>	PO on days 1-5
<i>Repeat treatment every 21 days.</i>		

<sup>a</sup> Methotrexate given as a 100-mg/m<sup>2</sup> IV bolus, then 300 mg/m<sup>2</sup> IV over 4 hours, followed 24 hours later by leucovorin, 15 mg PO, q6h for 6 doses.